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Si aucun titre n'est indiqué se referer à la description.)

ACC gene

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ACC gene

The present invention relates to a gene useful in a process to increase the microbial production of carotenoids.

5 The carotenoid astaxanthin is distributed in a wide variety of organisms such as animals, algae and microorganisms. It has a strong antioxidation property against reactive oxygen species. Astaxanthin is used as a coloring reagent, especially in the industry of farmed fish, such as salmon, because astaxanthin imparts distinctive orange-red coloration to the animals and contributes to consumer appeal in the marketplace.

10 One of the first steps in the carotenogenic pathway of, e.g. *Phaffia rhodozyma*, is the condensation of two molecules of acetyl-CoA. Acetyl-CoA is also the substrate for acetyl-CoA carboxylase, one of the enzymes involved in fatty acid biosynthesis.

In one aspect, the present invention provides a novel DNA fragment comprising a gene encoding the enzyme acetyl-CoA carboxylase.

15 More particularly, the present invention provides a DNA containing regulatory regions, such as promoter and terminator, as well as the open reading frame of acetyl-CoA carboxylase gene.

20 The present invention provides a DNA fragment encoding acetyl-CoA carboxylase in *P. rhodozyma*. The said DNA means a cDNA which contains only open reading frame flanked between the short fragments in its 5'- and 3'- untranslated region, and a genomic DNA which also contains its regulatory sequences such as its promoter and terminator which are necessary for the expression of the acetyl-CoA carboxylase gene in *P. rhodozyma*.

Accordingly, the present invention relates to a polynucleotide comprising a nucleic acid molecule selected from the group consisting of:

- (a) nucleic acid molecules encoding at least the mature form of the polypeptide depicted in SEQ ID NO:3;
- 5 (b) nucleic acid molecules comprising the coding sequence as depicted in SEQ ID NO:2;
- (c) nucleic acid molecules whose nucleotide sequence is degenerate as a result of the genetic code to a nucleotide sequence of (a) or (b);
- (d) nucleic acid molecules encoding a polypeptide derived from the polypeptide encoded by a polynucleotide of (a) to (c) by way of substitution, deletion and/or addition of one or
10 several amino acids of the amino acid sequence of the polypeptide encoded by a polynucleotide of (a) to (c);
- (e) nucleic acid molecules encoding a polypeptide derived from the polypeptide whose sequence has an identity of 56.3 % or more to the amino acid sequence of the polypeptide encoded by a nucleic acid molecule of (a) or (b);
- 15 (f) nucleic acid molecules comprising a fragment or an epitope-bearing portion of a polypeptide encoded by a nucleic acid molecule of any one of (a) to (e) and having acetyl-CoA carboxylase activity;
- (g) nucleic acid molecules comprising a polynucleotide having a sequence of a nucleic acid molecule amplified from *Phaffia* or *Xanthophylomyces* nucleic acid library using the
20 primers depicted in SEQ ID NO:4, 5, and 6;
- (h) nucleic acid molecules encoding a polypeptide having acetyl-CoA carboxylase activity, wherein said polypeptide is a fragment of a polypeptide encoded by any one of (a) to (g);
- (i) nucleic acid molecules comprising at least 15 nucleotides of a polynucleotide of any one of (a) to (d);
- 25 (j) nucleic acid molecules encoding a polypeptide having acetyl-CoA carboxylase activity, wherein said polypeptide is recognized by antibodies that have been raised against a polypeptide encoded by a nucleic acid molecule of any one of (a) to (h);
- (k) nucleic acid molecules obtainable by screening an appropriate library under stringent conditions with a probe having the sequence of the nucleic acid molecule of any one of (a)
30 to (j), and encoding a polypeptide having an acetyl-CoA carboxylase activity;
- (l) nucleic acid molecules whose complementary strand hybridizes under stringent conditions with a nucleic acid molecule of any one of (a) to (k), and encoding a polypeptide having acetyl-CoA carboxylase activity.

The terms "gene(s)", "polynucleotide", "nucleic acid sequence", "nucleotide sequence",
35 "DNA sequence" or "nucleic acid molecule(s)" as used herein refers to a polymeric form of

nucleotides of any length, either ribonucleotides or deoxyribonucleotides. This term refers only to the primary structure of the molecule.

Thus, this term includes double- and single- stranded DNA, and RNA. It also includes known types of modifications, for example, methylation, "caps" substitution of one or more of the naturally occurring nucleotides with an analog. Preferably, the DNA sequence of the invention comprises a coding sequence encoding the above-defined polypeptide.

A "coding sequence" is a nucleotide sequence which is transcribed into mRNA and/or translated into a polypeptide when placed under the control of appropriate regulatory sequences. The boundaries of the coding sequence are determined by a translation start codon at the 5'-terminus and a translation stop codon at the 3'-terminus. A coding sequence can include, but is not limited to mRNA, cDNA, recombinant nucleotide sequences or genomic DNA, while introns may be present as well under certain circumstances. SEQ ID:1 depicts the genomic DNA in which the intron sequence is inserted in the coding sequence for acetyl-CoA carboxylase gene from *P. rhodozyma*.

In general, the gene consists of several parts which have different functions from each other. In eukaryotes, genes which encode a corresponding protein, are transcribed to pre-mature messenger RNA (pre-mRNA) differing from the genes for ribosomal RNA (rRNA), small nuclear RNA (snRNA) and transfer RNA (tRNA). Although RNA polymerase II (PolII) plays a central role in this transcription event, PolII can not solely start transcription without *cis* element covering an upstream region containing a promoter and an upstream activation sequence (UAS), and a *trans*-acting protein factor. At first, a transcription initiation complex which consists of several basic protein components recognize the promoter sequence in the 5'-adjacent region of the gene to be expressed. In this event, some additional participants are required in the case of the gene which is expressed under some specific regulation, such as a heat shock response, or adaptation to a nutrition starvation, and so on. In such a case, a UAS is required to exist in the 5'-untranslated upstream region around the promoter sequence, and some positive or negative regulator proteins recognize and bind to the UAS. The strength of the binding of transcription initiation complex to the promoter sequence is affected by such a binding of the *trans*-acting factor around the promoter, and this enables the regulation of transcription activity.

After the activation of a transcription initiation complex by the phosphorylation, a transcription initiation complex initiates transcription from the transcription start site. Some parts of the transcription initiation complex are detached as an elongation complex from the promoter region to the 3' direction of the gene (this step is called as a promoter

clearance event) and the elongation complex continues the transcription until it reaches to a termination sequence that is located in the 3'-adjacent downstream region of the gene. Pre-mRNA thus generated is modified in nucleus by the addition of cap structure at the cap site which almost corresponds to the transcription start site, and by the addition of polyA stretches at the polyA signal which is located at the 3'-adjacent downstream region. Next, intron structures are removed from the coding region and exon parts are combined to yield an open reading frame whose sequence corresponds to the primary amino acid sequence of a corresponding protein. This modification in which a mature mRNA is generated is necessary for a stable gene expression. cDNA in general terms corresponds to the DNA sequence which is reverse-transcribed from this mature mRNA sequence. It can be synthesized by the reverse transcriptase derived from viral species by using a mature mRNA as a template, experimentally.

To express a gene which was derived from eukaryote, a procedure in which cDNA is cloned into an expression vector for *E. coli* is often used. This results from the fact that a specificity of intron structure varies among the organisms and an inability to recognize the intron sequence from other species. In fact, prokaryote has no intron structure in its own genetic background. Even in yeast, the genetic background is different between *Ascomycetes* to which *Saccharomyces cerevisiae* belongs and *Basidiomycetes* to which *P. rhodozyma* belongs, e.g. the intron structure of the actin gene from *P. rhodozyma* cannot be recognized nor spliced by the ascomycetous yeast, *S. cerevisiae*.

Intron structures of some kinds of the genes appear to be involved in the regulation of the expression of their genes. It might be important to use a genomic fragment which has its introns in a case of self-cloning of the gene of a interest whose intron structure involves such a regulation of its own gene expression.

To apply a genetic engineering method for a strain improvement study, it is necessary to study its genetic mechanism in the event such as transcription and translation. It is important to determine a genetic sequence such as its UAS, promoter, intron structure and terminator to study the genetic mechanism.

According to this invention, the gene encoding the acetyl-CoA carboxylase (ACC) gene from *P. rhodozyma* including its 5'- and 3'-adjacent regions as well as its intron structure was determined.

The invention further encompasses polynucleotides that differ from one of the nucleotide sequences shown in SEQ ID NO:2 (and portions thereof) due to degeneracy of the genetic

code and also encode an acetyl-CoA carboxylase as that encoded by the nucleotide sequences shown in SEQ ID NO:2. Further the polynucleotide of the invention has a nucleotide sequence encoding a protein having an amino acid sequence shown in SEQ ID NO:3. In a still further embodiment, the polynucleotide of the invention encodes a full length *P. rhodozyma* protein which is substantially homologous to an amino acid sequence of SEQ ID NO:3.

In addition, it will be appreciated by those skilled in the art that DNA sequence polymorphism that lead to changes in the amino acid sequences may exist within a population (e.g., the *P. rhodozyma* population). Such genetic polymorphism in the acetyl-CoA carboxylase gene may exist among individuals within a population due to natural variation.

As used herein, the terms "gene" and "recombinant gene" refer to nucleic acid molecules comprising an open reading frame encoding an acetyl-CoA carboxylase, preferably an acetyl-CoA carboxylase from *P. rhodozyma*.

Such natural variations can typically result in 1-5 % variance in the nucleotide sequence of the acetyl-CoA carboxylase gene. Any and all such nucleotide variations and resulting amino acid polymorphism in acetyl-CoA carboxylase that are the result of natural variation and that do not alter the functional activity of acetyl-CoA carboxylase are intended to be within the scope of the invention.

Polynucleotides corresponding to natural variants and non-*P. rhodozyma* homologues of the acetyl-CoA carboxylase cDNA of the invention can be isolated based on their homology to *P. rhodozyma* acetyl-CoA carboxylase polynucleotides disclosed herein using the polynucleotide of the invention, or a portion thereof, as a hybridization probe according to standard hybridization techniques under stringent hybridization conditions.

Accordingly, in another embodiment, a polynucleotide of the invention is at least 15 nucleotides in length. Preferably it hybridizes under stringent conditions to the nucleic acid molecule comprising a nucleotide sequence of the polynucleotide of the present invention, e.g. SEQ ID NO:2. In other embodiments, the nucleic acid is at least 20, 30, 50, 100, 250 or more nucleotides in length. The term "hybridizes under stringent conditions" is defined above and is intended to describe conditions for hybridization and washing under which nucleotide sequences at least 60% identical to each other typically remain hybridized to each other. Preferably, the conditions are such that sequences at least about 65% or 70%, more preferably at least about 75% or 80%, and even more preferably at least about 85%, 90% or 95% or more identical to each other typically remain hybridized to each other. Preferably, polynucleotide of the invention that hybridizes under stringent

conditions to a sequence of SEQ ID NO:2 corresponds to a naturally occurring nucleic acid molecule.

In the present invention, the polynucleotide sequence includes SEQ ID NO:2 and fragments thereof having polynucleotide sequences which hybridize to SEQ ID NO:2 under
5 stringent conditions which are sufficient to identify specific binding to SEQ ID NO:2. For example, any combination of the following hybridization and wash conditions may be used to achieve the required specific binding:

High Stringent Hybridization: 6X SSC, 0.5% SDS, 100 µg/ml denatured salmon sperm DNA, 50% formamide, incubate overnight with gentle rocking at 42°C.

10 High Stringent Wash: 1 wash in 2X SSC, 0.5% SDS at room temperature for 15 minutes, followed by another wash in 0.1X SSC, 0.5% SDS at room temperature for 15 minutes.

Low Stringent Hybridization: 6X SSC, 0.5% SDS, 100 µg/ml denatured salmon sperm DNA, 50% formamide, incubate overnight with gentle rocking at 37°C.

Low Stringent Wash: 1 wash in 0.1X SSC, 0.5% SDS at room temperature for 15 minutes.

15 Moderately stringent conditions may be obtained by varying the temperature at which the hybridization reaction occurs and/or the wash conditions as set forth above. In the present invention, it is preferred to use high stringent hybridization and wash conditions to define the antisense activity against acetyl-CoA carboxylase gene from *P. rhodozyma*.

The term "homology" means that the respective nucleic acid molecules or encoded proteins are functionally and/or structurally equivalent. The nucleic acid molecules that are
20 homologous to the nucleic acid molecules described above and that are derivatives of said nucleic acid molecules are, for example, variations of said nucleic acid molecules which represent modifications having the same biological function, in particular encoding proteins with the same or substantially the same biological function. They may be naturally occurring
25 variations, such as sequences from other plant varieties or species, or mutations. These mutations may occur naturally or may be obtained by mutagenesis techniques. The allelic variations may be naturally occurring allelic variants as well as synthetically produced or genetically engineered variants. Structural equivalents can, for example, be identified by testing the binding of said polypeptides to antibodies. Structural equivalents have similar
30 immunological characteristics, e.g. comprise similar epitopes.

As used herein, a "naturally-occurring" nucleic acid molecule refers to an RNA or DNA molecule having a nucleotide sequence that occurs in nature (e.g., encodes a natural protein). Preferably, the polynucleotide encodes a natural *P. rhodozyma* acetyl-CoA carboxylase.

acid or nucleic acid "homology" is equivalent to amino acid or nucleic acid "identity"). The percent homology between the two sequences is a function of the number of identical positions shared by the sequences (i.e., % homology = numbers of identical positions/total numbers of positions x 100). The homology can be determined by computer programs as

5 Blast 2.0 [Altschul, Nuc. Acid. Res., 25:3389-3402 (1997)]. In this invention, GENETYX-SV/RC software (Software Development Co., Ltd., Tokyo, Japan) is used by using its default algorithm as such homology analysis software. This software uses the Lipman-Pearson method for its analytic algorithm.

A nucleic acid molecule encoding an acetyl-CoA carboxylase homologous to a protein

10 with an amino acid sequence of SEQ ID NO:3 can be created by introducing one or more nucleotide substitutions, additions or deletions into a nucleotide sequence of the polynucleotide of the present invention, in particular of SEQ ID NO:2 such that one or more amino acid substitutions, additions or deletions are introduced into the encoded protein. Mutations can be introduced into the sequences of, e.g., SEQ ID NO:2 by

15 standard techniques, such as site-directed mutagenesis and PCR-mediated mutagenesis. Preferably, conservative amino acid substitutions are made at one or more predicted non-essential amino acid residues. A "conservative amino acid substitution" is one in which the amino acid residue is replaced with an amino acid residue having a similar side chain. Families of amino acid residues having similar side chains have been defined in the art.

20 These families include amino acids with basic side chains (e.g., lysine, arginine, histidine), acidic side chains (e.g., aspartic acid, glutamic acid), uncharged polar side chains (e.g., glycine, asparagine, glutamine, serine, threonine, tyrosine, cysteine), nonpolar side chains (e.g., alanine, valine, leucine, isoleucine, proline, phenylalanine, methionine, tryptophan), beta-branched side chains (e.g., threonine, valine, isoleucine) and aromatic side chains

25 (e.g., tyrosine, phenylalanine, tryptophan, histidine). Thus, a predicted nonessential amino acid residue in an acetyl-CoA carboxylase is preferably replaced with another amino acid residue from the same family. Alternatively, in another embodiment, mutations can be introduced randomly along all or part of an acetyl-CoA carboxylase coding sequence, such as by saturation mutagenesis, and the resultant mutants can be screened for an acetyl-

30 CoA carboxylase activity described herein to identify mutants that retain acetyl-CoA carboxylase activity. Following mutagenesis of one of the sequences of SEQ ID NO:2, the encoded protein can be expressed recombinantly and the activity of the protein can be determined using, for example, assays described herein.

A polynucleotide of the present invention, e.g., a nucleic acid molecule having a nucleotide

35 sequence of SEQ ID NO:2, or a portion thereof, can be isolated using standard molecular

Preferably, the polypeptide of the invention comprises one of the nucleotide sequences shown in SEQ ID NO:2. The sequence of SEQ ID NO:2 corresponds to the *P. rhodozyma* acetyl-CoA carboxylase cDNAs of the invention.

Further, the polynucleotide of the invention comprises a nucleic acid molecule which is a complement of one of the nucleotide sequences of above mentioned polynucleotides or a portion thereof. A nucleic acid molecule which is complementary to one of the nucleotide sequences shown in SEQ ID NO:2 is one which is sufficiently complementary to one of the nucleotide sequences shown in SEQ ID NO:2 such that it can hybridize to one of the nucleotide sequences shown in SEQ ID NO:2, thereby forming a stable duplex.

10 The polynucleotide of the invention comprises a nucleotide sequence which is at least about 60%, preferably at least about 65-70%, more preferably at least about 70-80%, 80-90%, or 90-95%, and even more preferably at least about 95%, 96%, 97%, 98%, 99% or more homologous to a nucleotide sequence shown in SEQ ID NO:2, or a portion thereof. The polynucleotide of the invention comprises a nucleotide sequence which hybridizes, e.g., hybridizes under stringent conditions as defined herein, to one of the nucleotide sequences shown in SEQ ID NO:2, or a portion thereof.

Moreover, the polynucleotide of the invention can comprise only a portion of the coding region of one of the sequences in SEQ ID NO:2, for example a fragment which can be used as a probe or primer or a fragment encoding a biologically active portion of an acetyl-CoA carboxylase. The nucleotide sequences determined from the cloning of the acetyl-CoA carboxylase gene from *P. rhodozyma* allows for the generation of probes and primers designed for use in identifying and/or cloning acetyl-CoA carboxylase homologues in other cell types and organisms. The probe/primer typically comprises a substantially purified oligonucleotide. The oligonucleotide typically comprises a region of nucleotide sequence that hybridizes under stringent conditions to at least about 12, 15 preferably about 20 or 25, more preferably about 40, 50 or 75 consecutive nucleotides of a sense strand of one of the sequences set forth, e.g., in SEQ ID NO: No:2; an anti-sense sequence of one of the sequences, e.g., set forth in SEQ ID NO:2, or naturally occurring mutants thereof. Primers based on a nucleotide of invention can be used in PCR reactions to clone acetyl-CoA carboxylase homologues. Probes based on the acetyl-CoA carboxylase nucleotide sequences can be used to detect transcripts or genomic sequences encoding the same or homologous proteins. The probe can further comprise a label group attached thereto, e.g., the label group can be a radioisotope, a fluorescent compound, an enzyme, or an enzyme co-factor. Such probes can be used as a part of a genomic marker test kit for identifying cells which express an acetyl-CoA carboxylase, such as by measuring a level of

an acetyl-CoA carboxylase-encoding nucleic acid molecule in a sample of cells, e.g., detecting acetyl-CoA carboxylase mRNA levels or determining whether a genomic acetyl-CoA carboxylase gene has been mutated or deleted.

The polynucleotide of the invention encodes a polypeptide or portion thereof which includes an amino acid sequence which is sufficiently homologous to an amino acid sequence of SEQ ID NO:3 such that the protein or portion thereof maintains an acetyl-CoA carboxylase activity, in particular an acetyl-CoA carboxylase activity as described in the examples in microorganisms or plants. As used herein, the language "sufficiently homologous" refers to proteins or portions thereof which have amino acid sequences which include a minimum number of identical or equivalent (e.g., an amino acid residue which has a similar side chain as an amino acid residue in one of the sequences of the polypeptide of the present invention amino acid residues to an amino acid sequence of SEQ ID NO:3 such that the protein or portion thereof has an acetyl-CoA carboxylase activity. Examples of an acetyl-CoA carboxylase activity are also described herein.

The protein is at least about 60-65%, preferably at least about 66-70%, and more preferably at least about 70-80%, 80-90%, 90-95%, and most preferably at least about 96%, 97%, 98%, 99% or more homologous to an entire amino acid sequence of SEQ ID NO:3.

Portions of proteins encoded by the acetyl-CoA carboxylase polynucleotide of the invention are preferably biologically active portions of one of the acetyl-CoA carboxylase.

As mentioned herein, the term "biologically active portion of acetyl-CoA carboxylase" is intended to include a portion, e.g., a domain/motif, that has acetyl-CoA carboxylase activity or has an immunological activity such that it binds to an antibody binding specifically to acetyl-CoA carboxylase. To determine whether an acetyl-CoA carboxylase or a biologically active portion thereof can participate in the metabolism an assay of enzymatic activity may be performed. Such assay methods are well known to those skilled in the art, as detailed in the Examples. Additional nucleic acid fragments encoding biologically active portions of an acetyl-CoA carboxylase can be prepared by isolating a portion of one of the sequences in SEQ ID NO:2, expressing the encoded portion of the acetyl-CoA carboxylase or peptide (e.g., by recombinant expression *in vitro*) and assessing the activity of the encoded portion of the acetyl-CoA carboxylase or peptide.

At first, a partial gene fragment was cloned containing a portion of the ACC gene by using the degenerate PCR method. Said degenerate PCR is a method to clone a gene of interest which has high homology of amino acid sequence to the known enzyme from other species

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which has the same or similar function. Degenerate primer, which is used as a primer in degenerate PCR, was designed by a reverse translation of the amino acid sequence to corresponding nucleotides ("degenerated"). In such a degenerate primer, a mixed primer which consists any of A, C, G or T, or a primer containing inosine at an ambiguity code is generally used. In this invention, such mixed primers were used for degenerate primers to clone above gene.

An entire gene containing its coding region with its intron as well as its regulation region such as a promoter or a terminator can be cloned from a chromosome by screening of a genomic library which is constructed in phage vector or plasmid vector in appropriate host, by using a partial DNA fragment obtained by degenerate PCR as described above as a probe after it was labeled. Generally, *E. coli* as a host strain and *E. coli* vector, a phage vector such as λ phage vector, or a plasmid vector such as pUC vector is often used in the construction of a library and a following genetic manipulation such as a sequencing, a restriction digestion, a ligation and the like. In this invention, an *EcoRI* genomic library of *P. rhodozyma* was constructed in the derivatives of λ vector, λ ZAPII. An insert size, what length of insert must be cloned, was determined by the Southern blot hybridization for the gene before construction of a library. In this invention, a DNA used for a probe was labeled with digoxigenin (DIG), a steroid hapten instead of conventional ^{32}P label, following the protocol which was prepared by the supplier (Boehringer-Mannheim, Mannheim, Germany). A genomic library constructed from the chromosome of *P. rhodozyma* was screened by using a DIG-labeled DNA fragment which had a portion of a gene of interest as a probe. Hybridized plaques were picked up and used for further study. When λ ZAPII (insert size was below 9kb) was used in the construction of the genomic library, in vivo excision protocol was conveniently used for the succeeding step of the cloning into the plasmid vector by using a derivative of single stranded M13 phage, Ex assist phage (Stratagene, La Jolla, USA). A plasmid DNA thus obtained was examined for sequencing.

In this invention, we used the automated fluorescent DNA sequencer, ALFred system (Pharmacia, Uppsala, Sweden) using an autocycle sequencing protocol in which the Taq DNA polymerase is employed in most cases of sequencing.

After the determination of the genomic sequence, a sequence of a coding region was used for a cloning of cDNA of corresponding gene. The PCR method was also exploited to clone cDNA fragment. The PCR primers whose sequences were identical to the sequence at the 5'- and 3'- end of the open reading frame (ORF) were synthesized with an addition of an appropriate restriction site, and PCR was performed by using those PCR primers. In

The present invention further relates to a vector in which the polynucleotide of the present invention is operatively linked to expression control sequences allowing expression in prokaryotic or eukaryotic host cells. The nature of such control sequences differs depending upon the host organism. In prokaryotes, control sequences generally include promoter, ribosomal binding site, and terminators. In eukaryotes, generally control sequences include promoters, terminators and, in some instances, enhancers, transactivators; or transcription factors.

The term "control sequence" is intended to include, at a minimum, components the presence of which are necessary for expression, and may also include additional advantageous components.

The term "operably linked" refers to a juxtaposition wherein the components so described are in a relationship permitting them to function in their intended manner. A control sequence "operably linked" to a coding sequence is ligated in such a way that expression of the coding sequence is achieved under conditions compatible with the control sequences. In case the control sequence is a promoter, it is obvious for a skilled person that double-stranded nucleic acid is used.

Regulatory sequences include those which direct constitutive expression of a nucleotide sequence in many types of host cell and those which direct expression of the nucleotide sequence only in certain host cells or under certain conditions. It will be appreciated by those skilled in the art that the design of the expression vector can depend on such factors as the choice of the host cell to be transformed, the level of expression of protein desired, etc. The expression vectors of the invention can be introduced into host cells to thereby produce proteins or peptides, including fusion proteins or peptides, encoded by polynucleotides as described herein.

The recombinant expression vectors of the invention can be designed for expression of acetyl-CoA carboxylase in prokaryotic or eukaryotic cells. For example, genes encoding the polynucleotide of the invention can be expressed in bacterial cells such as *E. coli*, insect cells (using baculovirus expression vectors), yeast and other fungal cells, algae, ciliates of the types: *Holotrichia*, *Peritrichia*, *Spirotrichia*, *Suctorina*, *Tetrahymena*, *Paramecium*, *Colpidium*, *Glaucoma*, *Platyophrya*, *Potomacus*, *Pseudocohnilembus*, *Euplotes*, *Engelmanniella*, and *Stylonychia*, especially *Stylonychia lemnae* with vectors following, a transformation method as described in WO9801572 and multicellular plant cells. Alternatively, the recombinant expression vector can be transcribed and translated *in vitro*, for example using T7 promoter regulatory sequences and T7 polymerase.

utilized in the bacterium chosen for expression, such as *E. coli*. Such alteration of nucleic acid sequences of the invention can be carried out by standard DNA synthesis techniques.

Further, the acetyl-CoA carboxylase vector can be a yeast expression vector. Examples of vectors for expression in yeast *S. cerevisiae* include pYepSec1, pMPa, pJRY88, and pYES2 (Invitrogen, San Diego, USA). Vectors and methods for the construction of vectors appropriate for use in other fungi, such as the filamentous fungi, are known to the skilled artisan:

Alternatively, the polynucleotide of the invention can be introduced in insect cells using baculovirus expression vectors. Baculovirus vectors available for expression of proteins in cultured insect cells (e.g., Sf9 cells) include the pAc series and the pVL series.

Alternatively, the polynucleotide of the invention is introduced in mammalian cells using a mammalian expression vector. Examples of mammalian expression vectors include pCDM8 and pMT2PC. When used in mammalian cells, the expression vector's control functions are often provided by viral regulatory elements. For example, commonly used promoters are derived from polyoma, Adenovirus 2, cytomegalovirus and Simian Virus 40.

The recombinant mammalian expression vector can be capable of directing expression of the nucleic acid preferentially in a particular cell type (e.g., tissue-specific regulatory elements are used to express the nucleic acid). Tissue-specific regulatory elements are known in the art. Non-limiting examples of suitable tissue-specific promoters include the albumin promoter (liver-specific), lymphoid-specific promoters, in particular promoters of T cell receptors and immunoglobulins, neuron-specific promoters (e.g., the neurofilament promoter), pancreas-specific promoters, and mammary gland-specific promoters (e.g., milk whey promoter; US 4,873, 316 and EP 264,166). Developmentally-regulated promoters are also encompassed, for example the murine hox promoters and the fetoprotein promoter.

Thus expressed ACC gene can be verified for its activity, e.g., by an enzyme assay method. Some experimental protocols are described in the literature. The following is the one of the methods which is used for the determination of acetyl-CoA carboxylase activity: Assays are performed by measuring the loss in acetyl-CoA and/or the production of malonyl-CoA at 5 min intervals for 20 min, using reverse phase HPLC. The rate of conversion of acetyl-CoA to malonyl-CoA is found to be linear for 20 min, and velocities are calculated by linear regression analysis of the malonyl-CoA concentration with respect to time. The

gene fragment would form a complex with a mature mRNA fragment of the objective gene *in vivo* and inhibit an efficient translation from mRNA, as a consequence.

An "antisense" nucleic acid molecule comprises a nucleotide sequence which is complementary to a "sense" nucleic acid molecule encoding a protein, e.g., complementary to the coding strand of a double-stranded cDNA molecule or complementary to a mRNA sequence. Accordingly, an antisense nucleic acid molecule can hydrogen bond to a sense nucleic acid molecule. The antisense nucleic acid molecule can be complementary to an entire acetyl-CoA carboxylase-coding strand, or to only a portion thereof. Accordingly, an antisense nucleic acid molecule can be antisense to a "coding region" of the coding strand of a nucleotide sequence encoding an acetyl-CoA carboxylase. The term "coding region" refers to the region of the nucleotide sequence comprising codons which are translated into amino acid residues. Further, the antisense nucleic acid molecule is antisense to a "noncoding region" of the coding strand of a nucleotide sequence encoding acetyl-CoA carboxylase. The term "noncoding region" refers to 5' and 3' sequences which flank the coding region that are not translated into a polypeptide (i.e., also referred to as 5' and 3' untranslated regions).

Given the coding strand sequences encoding acetyl-CoA carboxylase disclosed herein, antisense nucleic acid molecules of the invention can be designed according to the rules of Watson and Crick base pairing. The antisense nucleic acid molecule can be complementary to the entire coding region of acetyl-CoA carboxylase mRNA, but can also be an oligonucleotide which is antisense to only a portion of the coding or noncoding region of acetyl-CoA carboxylase mRNA. For example, the antisense oligonucleotide can be complementary to the region surrounding the translation start site of acetyl-CoA carboxylase mRNA. An antisense oligonucleotide can be, for example, about 5, 10, 15, 20, 25, 30, 35, 40, 45 or 50 nucleotides in length. An antisense nucleic acid molecule of the invention can be constructed using chemical synthesis and enzymatic ligation reactions using procedures known in the art. For example, an antisense nucleic acid molecule (e.g., an antisense oligonucleotide) can be chemically synthesized using naturally occurring nucleotides or variously modified nucleotides designed to increase the biological stability of the molecules or to increase the physical stability of the duplex formed between the antisense and sense nucleic acids, e.g., phosphorothioate derivatives and acridine substituted nucleotides can be used. Examples of modified nucleotides which can be used to generate the anti-sense nucleic acid include 5-fluorouracil, 5-bromouracil, 5-chlorouracil, 5-iodouracil, hypoxanthine, xanthine, 4-acetylcytosine, 5-(carboxyhydroxymethyl) uracil, 5-carboxymethylaminomethyl-2-thiouridine, 5-carboxymethylaminomethyluracil, dihydrouracil, beta-

In addition to naturally-occurring variants of the acetyl-CoA carboxylase sequence that may exist in the population, the skilled artisan will further appreciate that changes can be introduced by mutation into a nucleotide sequence of the polynucleotide encoding acetyl-CoA carboxylase, thereby leading to changes in the amino acid sequence of the encoded
5 acetyl-CoA carboxylase, without altering the functional ability of the acetyl-CoA carboxylase. For example, nucleotide substitutions leading to amino acid substitutions at "non-essential" amino acid residues can be made in a sequence of the polynucleotide encoding acetyl-CoA carboxylase, e.g. SEQ ID NO:2. A "non-essential" amino acid residue is a residue that can be altered from the wild-type sequence of one of the acetyl-CoA carboxylase
10 without altering the activity of said acetyl-CoA carboxylase, whereas an "essential" amino acid residue is required for acetyl-CoA carboxylase activity. Other amino acid residues, however, (e.g., those that are not conserved or only semi-conserved in the domain having acetyl-CoA carboxylase activity) may not be essential for activity and thus are likely to be amenable to alteration without altering acetyl-CoA carboxylase activity.

15 Accordingly, the invention relates to polynucleotides encoding acetyl-CoA carboxylase that contain changes in amino acid residues that are not essential for acetyl-CoA carboxylase activity. Such acetyl-CoA carboxylase differs in amino acid sequence from a sequence contained in SEQ ID NO:3 yet retain the acetyl-CoA carboxylase activity described herein. The polynucleotide can comprise a nucleotide sequence encoding a
20 polypeptide, wherein the polypeptide comprises an amino acid sequence at least about 60% identical to an amino acid sequence of SEQ ID NO:3 and has acetyl-CoA carboxylase activity. Preferably, the protein encoded by the nucleic acid molecule is at least about 60-65% identical to the sequence in SEQ ID NO:3, more preferably at least about 60-70% identical to one of the sequences in SEQ ID NO:3, even more preferably at least about 70-
25 80%, 80-90%, 90-95% homologous to the sequence in SEQ ID NO:3, and most preferably at least about 96%, 97%, 98%, or 99% identical to the sequence in SEQ ID NO:3.

To determine the percent homology of two amino acid sequences, (e.g., one of the sequence of SEQ ID NO:3 and a mutant form thereof) or of two nucleic acids, the sequences are aligned for optimal comparison purposes (e.g., gaps can be introduced in the sequence
30 of one protein or nucleic acid for optimal alignment with the other protein or nucleic acid). The amino acid residues or nucleotides at corresponding amino acid positions or nucleotide positions are then compared. When a position in one sequence (e.g., one of the sequences of SEQ ID NO:2 or 3) is occupied by the same amino acid residue or nucleotide as the corresponding position in the other sequence (e.g., a mutant form of the sequence
35 selected), then the molecules are homologous at that position (i.e., as used herein amino

biology techniques and the sequence information provided herein. For example, acetyl-CoA carboxylase cDNA can be isolated from a library using all or portion of one of the sequences of the polynucleotide of the present invention as a hybridization probe and standard hybridization techniques. Moreover, a polynucleotide encompassing all or a portion of one of the sequences of the polynucleotide of the present invention can be isolated by the polymerase chain reaction using oligonucleotide primers designed based upon this sequence (e.g., a nucleic acid molecule encompassing all or a portion of one of the sequences of polynucleotide of the present invention can be isolated by the polymerase chain reaction using oligonucleotide primers, e.g., of SEQ ID NO:4, 5, or 6, designed based upon this same sequence of polynucleotide of the present invention. For example, mRNA can be isolated from cells, e.g. *Phaffia* (e.g., by the guanidinium-thiocyanate extraction procedure of Chirgwin et al. and cDNA can be prepared using reverse transcriptase (e.g., Moloney MLV reverse transcriptase or AMV reverse transcriptase available from Promega (Madison, USA)). Synthetic oligonucleotide primers for polymerase chain reaction amplification can be designed based upon one of the nucleotide sequences shown in SEQ ID NO:2. A polynucleotide of the invention can be amplified using cDNA or, alternatively, genomic DNA, as a template and appropriate oligonucleotide primers according to standard PCR amplification techniques. The polynucleotide so amplified can be cloned into an appropriate vector and characterized by DNA sequence analysis. Furthermore, oligonucleotides corresponding to an acetyl-CoA carboxylase nucleotide sequence can be prepared by standard synthetic techniques, e.g., using an automated DNA synthesizer.

The terms "fragment", "fragment of a sequence" or "part of a sequence" means a truncated sequence of the original sequence referred to. The truncated sequence (nucleic acid or protein sequence) can vary widely in length; the minimum size being a sequence of sufficient size to provide a sequence with at least a comparable function and/or activity of the original sequence referred to, while the maximum size is not critical. In some applications, the maximum size usually is not substantially greater than that required to provide the desired activity and/or function(s) of the original sequence.

Typically, the truncated amino acid sequence will range from about 5 to about 60 amino acids in length. More typically, however, the sequence will be a maximum of about 50 amino acids in length, preferably a maximum of about 30 amino acids. It is usually desirable to select sequences of at least about 10, 12 or 15 amino acids, up to maximum of about 20 or 25 amino acids.

The term "epitope" relates to specific immunoreactive sites within an antigen, also known as antigenic determinants. These epitopes can be a linear array of monomers in a polymeric composition - such as amino acids in a protein - or consist of or comprise a more complex secondary or tertiary structure. Those of skill will recognize that all immunogens (i. e., substances capable of eliciting an immune response) are antigens; however, some antigen, such as haptens, are not immunogens but may be made immunogenic by coupling to a carrier molecule. The term "antigen" includes references to a substance to which an antibody can be generated and/or to which the antibody is specifically immunoreactive.

10 The term "one or several amino acids" relates to at least one amino acid but not more than that number of amino acids which would result in a homology of below 60% identity. Preferably, the identity is more than 70% or 80%, more preferred are 85%, 90% or 95%, even more preferred are 96%, 97%, 98%, or 99% identity.

The term "acetyl-CoA carboxylase" or "acetyl-CoA carboxylase activity" relates to enzymatic activities of a polypeptide as described below or which can be determined in enzyme assay method. Furthermore, polypeptides that are inactive in an assay herein but are recognized by an antibody specifically binding to acetyl-CoA carboxylase, i.e., having one or more acetyl-CoA carboxylase epitopes, are also comprised under the term "acetyl-CoA carboxylase". In these cases activity refers to their immunological activity.

20 The terms "polynucleotide" and "nucleic acid molecule" also relate to "isolated" polynucleotides or nucleic acids molecules. An "isolated" nucleic acid molecule is one which is separated from other nucleic acid molecules which are present in the natural source of the nucleic acid. Preferably, an "isolated" nucleic acid is free of sequences which naturally flank the nucleic acid (i.e., sequences located at the 5' and 3' ends of the nucleic acid) in the genomic DNA of the organism from which the nucleic acid is derived.

For example, in various embodiments, the PNO polynucleotide can contain less than about 5 kb, 4kb, 3kb, 2kb, 1 kb, 0.5 kb or 0.1 kb of nucleotide sequences which naturally flank the nucleic acid molecule in genomic DNA of the cell from which the nucleic acid is derived (e.g., a *Phaffia* cell). Moreover, the polynucleotides of the present invention, in particular an "isolated" nucleic acid molecule, such as a cDNA molecule, can be substantially free of other cellular material, or culture medium when produced by recombinant techniques, or chemical precursors or other chemicals when chemically synthesized.

D-galactosylqucosine, inosine, N6-isopentenyladenine, 1-methylguanine, 1-methylinosine, 2,2-dimethylguanine, 2-methyladenine, 2-methylguanine, 3-methylcytosine, 5-methylcytosine, N6-adenine, 7-methylguanine, 5-methylaminomethyluracil, 5-methoxyaminomethyl-2-thiouracil, beta-D-mannosylqucosine, 5'-methoxycarboxymethyluracil, 5-methoxyuracil, 2-methylthio-N6-isopentenyladenine, uracil-5-oxyacetic acid (v), wybutoxosine, pseudouracil, queosine, 2-thiocytosine, 5-methyl-2-thiouracil, 2-thiouracil, 4-thiouracil, 5-methyluracil, uracil-5-oxyacetic acid methylester, uracil-5-oxyacetic acid (v), 5-methyl-2-thiouracil, 3-(3-amino-3-N-2-carboxypropyl) uracil, (acp3)w, and 2,6-diaminopurine. Alternatively, the antisense nucleic acid can be produced biologically using an expression vector into which a polynucleotide has been subcloned in an antisense orientation (i.e., RNA transcribed from the inserted polynucleotide will be of an antisense orientation to a target polynucleotide of interest, described further in the following subsection).

The antisense nucleic acid molecules of the invention are typically administered to a cell or generated in situ such that they hybridize with or bind to cellular mRNA and/or genomic DNA encoding an acetyl-CoA carboxylase to thereby inhibit expression of the protein, e.g., by inhibiting transcription and/or translation. The hybridization can be by conventional nucleotide complementarity to form a stable duplex, or, for example, in the case of an antisense nucleic acid molecule which binds to DNA duplexes, through specific interactions in the major groove of the double helix. The anti-sense molecule can be modified such that it specifically binds to a receptor or an antigen expressed on a selected cell surface, e.g., by linking the antisense nucleic acid molecule to a peptide or an antibody which binds to a cell surface receptor or antigen. The antisense nucleic acid molecule can also be delivered to cells using the vectors described herein. To achieve sufficient intracellular concentrations of the antisense molecules, vector constructs in which the antisense nucleic acid molecule is placed under the control of a strong prokaryotic, viral, or eukaryotic including plant promoters are preferred.

The antisense nucleic acid molecule of the invention may, e.g., be an α -anomeric nucleic acid molecule. An α -anomeric nucleic acid molecule forms specific double-stranded hybrids with complementary RNA in which, contrary to the usual β -units, the strands run parallel to each other. The antisense nucleic acid molecule can also comprise a 2'-o-methylribonucleotide or a chimeric RNA-DNA analogue.

Further the antisense nucleic acid molecule of the invention can be a ribozyme. Ribozymes are catalytic RNA molecules with ribonuclease activity which are capable of cleaving a

single-stranded nucleic acid, such as a mRNA, to which they have a complementary region. Thus, ribozymes (e.g., hammerhead ribozymes) can be used to catalytically cleave acetyl-CoA carboxylase mRNA transcripts to thereby inhibit translation of mRNA. A ribozyme having specificity for an acetyl-CoA carboxylase-encoding nucleic acid molecule
5 can be designed based upon the nucleotide sequence of an acetyl-CoA carboxylase cDNA disclosed herein or on the basis of a heterologous sequence to be isolated according to methods taught in this invention. For example, a derivative of a Tetrahymena L-19 IVS RNA can be constructed in which the nucleotide sequence of the active site is complementary to the nucleotide sequence to be cleaved in an encoding mRNA (see, e.g., US
10 4,987,071 and US 5,116,742). Alternatively, acetyl-CoA carboxylase mRNA can be used to select a catalytic RNA having a specific ribonuclease activity from a pool of RNA molecules.

The application of the antisense method to construct a carotenoid overproducing strain from *P. rhodozyma* is disclosed in EP 1,158,051.

15 In one embodiment the present invention relates to a method of making a recombinant host cell comprising introducing the vector or the polynucleotide of the present invention into a host cell.

Vector DNA can be introduced into prokaryotic or eukaryotic cells via conventional transformation or transfection techniques. As used herein, the terms "transformation" and
20 "transfection", conjugation and transduction are intended to refer to a variety of art-recognized techniques for introducing foreign nucleic acid (e.g., DNA) into a host cell, including calcium phosphate or calcium chloride co-precipitation, DEAE-dextran-mediated transfection, lipofection, natural competence, chemical-mediated transfer, or electroporation. Suitable methods for transforming or transfecting host cells including
25 plant cells are known to the skilled artisan.

For stable transfection of mammalian cells, only a small fraction of cells may integrate the foreign DNA into their genome, depending upon the expression vector and transfection technique used. In order to identify and select these integrants, a gene that encodes a selectable marker (e.g., resistance to antibiotics) is generally introduced into the host cells
30 along with the gene of interest. Preferred selectable markers include those which confer resistance to drugs, such as G418, hygromycin and methotrexate. Nucleic acid encoding a selectable marker can be introduced into a host cell on the same vector as that encoding the polypeptide of the present invention or can be introduced on a separate vector. Cells stably transfected with the introduced nucleic acid can be identified by, for example, drug

this invention, a cDNA pool was used as a template in this PCR cloning of cDNA. The said cDNA pool consists of various cDNA species which were synthesized *in vitro* by the viral reverse transcriptase and Taq polymerase (CapFinder Kit manufactured by Clontech, Palo Alto, U.S.A.) by using the mRNA obtained from *P. rhodozyma* as a template. cDNA of interest thus obtained was confirmed in its sequence. Furthermore, cDNA thus obtained was used for a confirmation of its enzyme activity after the cloning of the cDNA fragment into an expression vector which functions in *E. coli* under the strong promoter activity such as the *lac* or T7 expression system.

In another embodiment, the present invention relates to a method for making a recombinant vector comprising inserting a polynucleotide of the invention into a vector.

Further, the present invention relates to a recombinant vector containing the polynucleotide of the invention or produced by said method of the invention.

As used herein, the term "vector" refers to a nucleic acid molecule capable of transporting a polynucleotide to which it has been linked. One type of vector is a "plasmid", which refers to a circular double stranded DNA loop into which additional DNA segments can be ligated. Another type of vector is a viral vector, wherein additional DNA or PNA segments can be ligated into the viral genome. Certain vectors are capable of autonomous replication in a host cell into which they are introduced (e.g., bacterial vectors having a bacterial origin of replication and episomal mammalian vectors). Other vectors (e.g., non-episomal mammalian vectors) are integrated into the genome of a host cell upon introduction into the host cell, and thereby are replicated along with the host genome. Moreover, certain vectors are capable of directing the expression of genes to which they are operatively linked. Such vectors are referred to herein as "expression vectors". In general, expression vectors of utility in recombinant DNA techniques are often in the form of plasmids. In the present specification, "plasmid" and "vector" can be used interchangeably as the plasmid is the most commonly used form of vector. However, the invention is intended to include such other forms of expression vectors, such as viral vectors (e.g., replication defective retroviruses, adenoviruses and adeno-associated viruses), which serve equivalent functions.

The present invention also relates to cosmids, viruses, bacteriophages and other vectors used conventionally in genetic engineering that contain a nucleic acid molecule according to the invention. Methods which are well known to those skilled in the art can be used to construct various plasmids and vectors. Alternatively, the nucleic acid molecules and vectors of the invention can be reconstituted into liposomes for delivery to target cells.

Expression of proteins in prokaryotes is most often carried out with vectors containing constitutive or inducible promoters directing the expression of either fusion or non-fusion proteins. Fusion vectors add a number of amino acids to a protein encoded therein, usually to the amino terminus of the recombinant protein but also to the C-terminus or fused within suitable regions in the proteins. Such fusion vectors typically serve three purposes: 1) to increase expression of recombinant protein; 2) to increase the solubility of the recombinant protein; and 3) to aid in the purification of the recombinant protein by acting as a ligand in affinity purification. Often, in fusion expression vectors, a proteolytic cleavage site is introduced at the junction of the fusion moiety and the recombinant protein to enable separation of the recombinant protein from the fusion moiety subsequent to purification of the fusion protein. Such enzymes, and their cognate recognition sequences, include Factor Xa, thrombin and enterokinase.

Typical fusion expression vectors include pGEX (Pharmacia Biotech Inc.), pMAL (New England Biolabs, Beverly, USA) and pRIT5 (Pharmacia, Piscataway, USA) which fuse glutathione S-transferase (GST), maltose E binding protein, or protein A, respectively, to the target recombinant protein. In one embodiment, the coding sequence of the polypeptide encoded by the polynucleotide of the present invention is cloned into a pGEX expression vector to create a vector encoding a fusion protein comprising, from the N-terminus to the C-terminus, GST-thrombin cleavage site-X-protein. The fusion protein can be purified by affinity chromatography using glutathione-agarose resin, e.g. recombinant acetyl-CoA carboxylase unfused to GST can be recovered by cleavage of the fusion protein with thrombin.

Examples of suitable inducible non-fusion *E. coli* expression vectors include pTrc and pET 11d. Target gene expression from the pTrc vector relies on host RNA polymerase transcription from a hybrid trp-lac fusion promoter. Target gene expression from the pET 11d vector relies on transcription from a T7 *g_{nl}*-lac fusion promoter mediated by a coexpressed viral RNA polymerase (T7 *g_{nl}*). This viral polymerase is supplied by host strains BL21(DE3) or HMS174(DE3) from a resident λ prophage harboring a T7 *g_{nl}* gene under the transcriptional control of the *lacUV 5* promoter.

One strategy to maximize recombinant protein expression is to express the protein in host bacteria with an impaired capacity to proteolytically cleave the recombinant protein. Another strategy is to alter the nucleic acid sequence of the nucleic acid to be inserted into an expression vector so that the individual codons for each amino acid are those preferentially

reaction mixture contained 50 mM Tris, pH 7.5, 6 μ M acetyl-CoA, 2 mM ATP, 7 mM KHCO_3 , 8 mM MgCl_2 , 1 mM dithiothreitol, and 1 mg/ml bovine serum albumin. Enzyme is preincubated (30 min, 25°C) with bovine serum albumin (2 mg/ml) and potassium citrate (10 mM). Reactions are initiated by transferring 50 μ l of preincubated enzyme to
5 the reaction mixture (final volume 200 μ l) and incubated for 5-20 min at 25°C. Reactions are terminated by addition of 50 μ l 10% perchloric acid. Following termination of the reaction, the samples are centrifuged (3 min, 10,000 \times g) and analyzed by HPLC. A mobile phase of 10 mM KH_2PO_4 , pH 6.7 (solvent A), and MeOH (solvent B) is used. The flow rate is 1.0 ml/min, and the gradient is as follows: hold at 100% solvent A for 1 min
10 followed by a linear gradient to 30% solvent B over the next 5 min, then hold at 30% solvent B for 5 min. Using this method the retention times were 7.5 and 9.0 min for malonyl-CoA and acetyl-CoA, respectively. When an expression vector for *S. cerevisiae* is used, a complementation analysis can be conveniently exploited by using conditional acetyl-CoA carboxylase null mutant strain derived from *S. cerevisiae* as a host strain for its
15 confirmation of activity.

Succeeding to the confirmation of the enzyme activity, an expressed protein would be purified and used for raising the antibody against the purified enzyme. Antibody thus prepared would be used for a characterization of the expression of the corresponding enzyme in a strain improvement study, an optimization study of the culture condition,
20 and the like.

In a further embodiment, the present invention relates to an antibody that binds specifically to the polypeptide of the present invention or parts, i.e. specific fragments or epitopes of such a protein.

The antibodies of the invention can be used to identify and isolate other acetyl-CoA carboxylase and genes. These antibodies can be monoclonal antibodies, polyclonal antibodies
25 or synthetic antibodies as well as fragments of antibodies, such as Fab, Fv or scFv fragments etc. Monoclonal antibodies can be prepared, for example, by the techniques as originally described by Kohler and Milstein, which comprise the fusion of mouse myeloma cells to spleen cells derived from immunized mammals.

30 Furthermore, antibodies or fragments thereof to the aforementioned peptides can be obtained by using methods known to the skilled person. These antibodies can be used, for example, for the immunoprecipitation and immunolocalization of proteins according to the invention as well as for the monitoring of the synthesis of such proteins, for example, in recombinant organisms, and for the identification of compounds interacting with the

protein according to the invention. For example, surface plasmon resonance as employed in the BLAcore system can be used to increase the efficiency of phage antibodies selections, yielding a high increment of affinity from a single library of phage antibodies which bind to an epitope of the protein of the invention. In many cases, the binding phenomenon of
5 antibodies to antigens is equivalent to other ligand/anti-ligand binding.

In this invention, the gene fragment for acetyl-CoA carboxylase was cloned from *P. rhodo-*
zyrna with a purpose to decrease its expression level in *P. rhodozyrna* by genetic method
using the cloned gene fragment.

To decrease a gene expression with genetic methods, some strategies can be employed, one
10 of which is a gene-disruption method. In this method, a partial fragment of the objective
gene to be disrupted is ligated to a drug resistant cassette on the integration vector which
can not replicate in the host organism. A drug resistance gene which encodes the enzyme
that enables the host to survive in the presence of a toxic antibiotic is often used for the
selectable marker. G418 resistance gene harbored in pGB-Ph9 (Wery *et al.* (Gene, 184, 89-
15 97, 1997)) is an example of a drug resistance gene which functions in *P. rhodozyrna*.

Nutrition complementation marker can be also used in the host which has an appropriate
auxotrophy marker. *P. rhodozyrna* ATCC24221 strain that requires cytidine for its growth
is one example of the auxotroph. By using CTP synthetase as donor DNA for ATCC24221,
a host vector system using a nutrition complementation can be established.

20 After the transformation of the host organisms and recombination between the objective
gene fragment on the vector and its corresponding gene fragment on the chromosome of
the host organisms, the integration vector is integrated onto the host chromosome by
single cross recombination. As a result of this recombination, the drug resistant cassette
would be inserted in the objective gene whose translated product is only synthesized in its
25 truncated form which does not have its enzymatic function. In a similar manner, two
parts of the objective gene were also used for gene disruption study in which the drug
resistant gene can be inserted between such two partial fragments of the objective genes on
the integration vector. In the case of this type of vector, double recombination event
between the gene fragments harbored on the integration vector and the corresponding
30 gene fragments on the chromosome of the host are expected. Although frequency of this
double crossing-over recombination is lower than single cross recombination, null
phenotype of the objective gene by the double cross recombination is more stable than by
the single cross recombination.

On the other hand, this strategy has difficulty in the case of the gene whose function is essential and disruption is lethal for the host organism such as acetyl-CoA carboxylase gene. The function of acetyl-CoA carboxylase is indispensable for the host survival other than the biosynthesis of fatty acid. From such a viewpoint, it seemed to be difficult to construct the acetyl-CoA carboxylase disruptant from *P. rhodozyma* by this gene disruption method.

In such a case, other strategies can be applied to decrease (not to disrupt) a gene expression, one of which is a conventional mutagenesis to screen the mutant whose expression for acetyl-CoA carboxylase is decreased. In this method, an appropriate recombinant in which an appropriate reporter gene is fused to the promoter region of acetyl-CoA carboxylase gene from the host organism is mutated and mutants which show a weaker activity of reporter gene product can be screened. In such mutants, it is expected that their expression of acetyl-CoA carboxylase activity decreased by the mutation lying in the promoter region of reporter gene or *trans*-acting region which might affect the expression of acetyl-CoA carboxylase gene other than the mutation lying in the promoter gene itself. In the case of mutation occurring at the promoter region of the reporter fusion, such mutation can be isolated by the sequence of the corresponding region. Thus isolated mutation can be introduced in a variety of carotenoids, especially astaxanthin producing mutants derived from *P. rhodozyma* by a recombination between the original promoter for acetyl-CoA carboxylase gene on the chromosome and the mutated promoter fragment. To exclude mutations occurring at a *trans*-acting region, a mutation can also be induced by an *in vitro* mutagenesis of a *cis* element in the promoter region. In this approach, a gene cassette, containing a reporter gene which is fused to a promoter region derived from a gene of interest at its 5'-end and a terminator region from a gene of interest at its 3'-end, is mutagenized and then introduced into *P. rhodozyma*. By detecting the difference of the activity of the reporter gene, an effective mutation can be screened. Such a mutation can be introduced in the sequence of the native promoter region on the chromosome by the same method as the case of an *in vivo* mutation approach. But, these methods have some drawbacks to have some time-consuming process.

Another strategy to decrease a gene expression is an antisense method. This method is frequently applied to decrease the gene expression even when teleomorphic organisms such as *P. rhodozyma* are used as host organisms, to which the mutation and gene disruption method is usually difficult to be applied. The anti-sense method is a method to decrease an expression of gene of interest by introducing an artificial gene fragment, whose sequence is complementary to cDNA fragment of the gene of interest. Such an anti-sense

extrachromosomally. In this respect, it is also to be understood that the nucleic acid molecule of the invention can be used to restore or create a mutant gene via homologous recombination.

Accordingly, in another embodiment the present invention relates to a host cell genetically engineered with the polynucleotide of the invention or the vector of the invention.

The terms "host cell" and "recombinant host cell" are used interchangeably herein. It is understood that such terms refer not only to the particular subject cell but to the progeny or potential progeny of such a cell. Because certain modifications may occur in succeeding generations due to either mutation or environmental influences, such progeny may not, in fact, be identical to the parent cell, but are still included within the scope of the term as used herein.

For example, a polynucleotide of the present invention can be introduced in bacterial cells as well as insect cells, fungal cells or mammalian cells (such as Chinese hamster ovary cells (CHO) or COS cells), algae, ciliates, plant cells, fungi or other microorganisms like *E. coli*. Other suitable host cells are known to those skilled in the art. Preferred are *E. coli*, baculovirus, *Agrobacterium* or fungal cells are, for example, those of the genus *Saccharomyces*, e.g. those of the species *S. cerevisiae* or *P. rhodozyma* (*Xanthophylomyces dendrorhous*).

In addition, in one embodiment, the present invention relates to a method for the production of fungal transformants comprising the introduction of the polynucleotide or the vector of the present invention into the genome of said fungal cell.

For the expression of the nucleic acid molecules according to the invention in sense or antisense orientation in plant cells, the molecules are placed under the control of regulatory elements which ensure the expression in fungal cells. These regulatory elements may be heterologous or homologous with respect to the nucleic acid molecule to be expressed as well with respect to the fungal species to be transformed.

In general, such regulatory elements comprise a promoter active in fungal cells. To obtain constitutive expression in fungal cells, preferably constitutive promoters are used, e.g., the glyceraldehyde-3-dehydrogenase promoter derived from *P. rhodozyma* (WO 97/23,633). Inducible promoters may be used in order to be able to exactly control expression. An example for inducible promoters is the promoter of genes encoding heat shock proteins. Also an amylase gene promoter which is a candidate for such inducible promoters has been described (EP 1,035,206). The regulatory elements may further comprise transcriptional and/or translational enhancers functional in fungal cells. Furthermore, the regula-

selection (e.g., cells that have incorporated the selectable marker gene will survive, while the other cells die).

To create a homologous recombinant microorganism, a vector is prepared which contains at least a portion of the polynucleotide of the present invention into which a deletion, addition or substitution has been introduced to thereby alter, e.g., functionally disrupt, the acetyl-CoA carboxylase gene. Preferably, this acetyl-CoA carboxylase gene is a *P. rhodozyma* acetyl-CoA carboxylase gene, but it can be a homologue from a related or different source. Alternatively, the vector can be designed such that, upon homologous recombination, the endogenous acetyl-CoA carboxylase gene is mutated or otherwise altered but still encodes a functional protein (e.g., the upstream regulatory region can be altered to thereby alter the expression of the endogenous acetyl-CoA carboxylase). To create a point mutation via homologous recombination also DNA-RNA hybrids can be used known as chimeraplasty known from Cole-Strauss *et al.*, Nucl. Aci. Res., 27, 5, 1323-1330, 1999 and Kmiec, Gene therapy., American Scientist. 87, 3, 240-247. 1999.

- 15 The vector is introduced into a cell and cells in which the introduced polynucleotide gene has homologously recombined with the endogenous acetyl-CoA carboxylase gene are selected, using art-known techniques.

Further host cells can be produced which contain selection systems which allow for regulated expression of the introduced gene. For example, inclusion of the polynucleotide of the invention on a vector placing it under control of the lac operon permits expression of the polynucleotide only in the presence of IPTG. Such regulatory systems are well known in the art.

Preferably, the introduced nucleic acid molecule is foreign to the host cell.

By "foreign" it is meant that the nucleic acid molecule is either heterologous with, respect to the host cell, this means derived from a cell or organism with a different genomic background, or is homologous with respect to the host cell but located in a different genomic environment than the naturally occurring counterpart of said nucleic acid molecule. This means that, if the nucleic acid molecule is homologous with respect to the host cell, it is not located in its natural location in the genome of said host cell, in particular it is surrounded by different genes. In this case the nucleic acid molecule may be either under the control of its own promoter or under the control of a heterologous promoter. The vector or nucleic acid molecule according to the invention which is present in the host cell may either be integrated into the genome of the host cell or it may be maintained in some form

tory elements may include transcription termination signals, such as a poly-A signal, which lead to the addition of a poly A tail to the transcript which may improve its stability.

Methods for the introduction of foreign DNA into fungal cells are also well known in the art. These include, for example, transformation with the LiCl method, the fusion of proto-
5 plasts, electroporation, biolistic methods like particle bombardment other methods known in the art. Methods for the transformation using biolistic methods are well known to the person skilled in the art.

The term "transformation" as used herein, refers to the transfer of an exogenous polynucleotide into a host cell, irrespective of the method used for the transfer. The poly-
10 nucleotide may be transiently or stably introduced into the host cell and may be maintained non- integrated, for example, as a plasmid or as chimeric links, or alternatively, may be integrated into the host genome.

In general, the fungi which can be modified according to the invention and which either show overexpression of a protein according to the invention or a reduction of the synthesis
15 of such a protein can be derived from any desired fungal species.

Further, in one embodiment, the present invention relates to a fungal cell comprising the polynucleotide the vector or obtainable by the method of the present invention.

Thus, the present invention relates also to transgenic fungal cells which contain (preferably stably integrated into the genome) a polynucleotide according to the invention linked to
20 regulatory elements which allow expression of the polynucleotide in fungal cells and wherein the polynucleotide is foreign to the transformed fungal cell. For the meaning of foreign; see supra.

Thus, the present invention also relates to transformed fungal cells according to the invention.

25 Accordingly, due to the altered expression of acetyl-CoA carboxylase, cells metabolic pathways are modulated in yield production, and/or efficiency of production.

The terms "production" or "productivity" are art-recognized and include the concentration of the fermentation product (for example fatty acids, carotenoids, (poly)saccharides, lipids, vitamins, isoprenoids, wax esters, and/or polymers like polyhydroxyalkanoates
30 and/or its metabolism products or further desired fine chemical as mentioned herein) formed within a given time and a given fermentation volume (e.g., kg product per hour per liter).

The term "efficiency" of production includes the time required for a particular level of production to be achieved (for example, how long it takes for the cell to attain a particular rate of output of a said altered yield, in particular, into carotenoids, (poly)saccharides, lipids, vitamins, isoprenoids etc.).

- 5 The term "yield" or "product/carbon yield" is art-recognized and includes the efficiency of the conversion of the carbon source into the product (i.e. acetyl CoA, fatty acids, vitamins, carotenoids, isoprenoids, lipids etc. and/or further compounds as defined above and which biosynthesis is based on said products). This is generally written as, for example, kg product per kg carbon source. By increasing the yield or production of the compound, the
- 10 quantity of recovered molecules, or of useful recovered molecules of that compound in a given amount of culture over a given amount of time is increased.

- The terms "biosynthesis" (which is used synonymously for "synthesis" of "biological production" in cells, tissues plants, etc.) or a "biosynthetic pathway" are art-recognized and include the synthesis of a compound, preferably an organic compound, by a cell from
- 15 intermediate compounds in what may be a multistep and highly regulated process.

- The language "metabolism" is art-recognized and includes the totality of the biochemical reactions that take place in an organism. The metabolism of a particular compound, then, (e.g., the metabolism of acetyl CoA, a fatty acid, hexose, isoprenoid, vitamin, carotenoid, lipid etc.) comprises the overall biosynthetic, modification, and degradation pathways in
- 20 the cell related to this compound.

- Such a genetically engineered *P. rhodozyma* would be cultivated in an appropriate medium and evaluated in its productivity of carotenoids, especially astaxanthin. A hyper producer of astaxanthin thus selected would be confirmed in view of the relationship between its productivity and the level of gene or protein expression which is introduced by such a
- 25 genetic engineering method.

The present invention is further illustrated with Examples described below.

The following materials and methods employed in the Examples are described below:

Strains

- P. rhodozyma* ATCC96594 (re-deposited under the accession No. ATCC 74438 on April 8,
- 30 1998 pursuant to the Budapest Treaty)

E. coli DH5 α : F, ϕ 80d, *lacZ*AM15, Δ (*lacZYA-argF*)U169, *hsd* (r_K^- ; m_K^+), *recA1*, *endA1*, *deoR*, *thi-1*, *supE44*, *gyrA96*, *relA1* (Toyobo, Osaka, Japan)

Isolation of total RNA from *P. rhodozyma* was performed with the phenol method by using Isogen (Nippon Gene, Toyama, Japan). mRNA was purified from total RNA thus obtained by using mRNA separation kit (Clontech). cDNA was synthesized by using CapFinder cDNA construction kit (Clontech).

5 *In vitro* packaging was performed by using Gigapack III gold packaging extract (Stratagene).

The polymerase chain reaction (PCR) was performed with the thermal cycler from Perkin Elmer model 2400. Each PCR condition is described in examples. PCR primers were purchased from a commercial supplier. Fluorescent DNA primers for DNA sequencing were
10 purchased from Pharmacia. DNA sequencing was performed with the automated fluorescent DNA sequencer (ALFred, Pharmacia).

Competent cells of DH5 α were purchased from Toyobo (Japan).

Example 1: Isolation of mRNA from *P. rhodozyma* and construction of cDNA library

To construct cDNA library of *P. rhodozyma*, total RNA was isolated by phenol extraction
15 method right after the cell disruption and the mRNA from *P. rhodozyma* ATCC96594 strain was purified by using mRNA separation kit (Clontech).

At first, Cells of ATCC96594 strain from 10 ml of two-day-culture in YPD medium were harvested by centrifugation (1500 x g for 10 min.) and washed once with extraction buffer (10 mM Na-citrate / HCl (pH 6.2) containing 0.7 M KCl). After suspending in 2.5 ml of
20 extraction buffer, the cells were disrupted by French press homogenizer (Ohtake Works Corp., Tokyo, Japan) at 1500 kgf/cm² and immediately mixed with two times of volume of isogen (Nippon gene) according to the method specified by the manufacturer. In this step, 400 μ g of total RNA was recovered.

Then, this total RNA was purified by using mRNA separation kit (Clontech) according to
25 the method specified by the manufacturer. Finally, 16 μ g of mRNA from *P. rhodozyma* ATCC96594 strain was obtained.

To construct cDNA library, CapFinder PCR cDNA construction kit (Clontech) was used according to the method specified by the manufacturer. One μ g of purified mRNA was applied for a first strand synthesis followed by PCR amplification. After this amplification
30 by PCR, 1 mg of cDNA pool was obtained.

Example 2: Cloning of a partial ACC (acetyl-CoA carboxylase) gene from *P. rhodozyma*

E. coli XL1-Blue MRF': $\Delta(mcrA)183$, $\Delta(mcrCB-hsdSMR-mrr)173$, *endA1*, *supE44*, *thi-1*, *recA1*, *gyrA96*, *relA1*, *lac* [*F'* *proAB*, *lacIqZ* Δ M15, Tn10 (*tet*^r)] (Stratagene, La Jolla, USA)
E. coli SOLR: *e14-(mcrA)*, $\Delta(mcrCB-hsdSMR-mrr)171$, *sbcC*, *recB*, *recJ*, *umuC* :: Tn5(*kan*^r), *uvrC*, *lac*, *gyrA96*, *relA1*, *thi-1*, *endA1*, Δ R, [*F'* *proAB*, *lacIqZ* Δ M15] Su-(nonsuppressing)

5 (Stratagene)

E. coli TOP10: *F*-, *mcrA*, $\Delta mrr-hsdRMS-mcrBC$, $\phi 80$, $\Delta lacZ$ M15, $\Delta lacX74$, *recA1*, *deoR*, *araD139*, (*ara-leu*)7697, *galU*, *galK*, *rpsL* (*Str*^r), *endA1*, *nupG* (Invitrogen, Carlsbad, USA)

Vectors

λ ZAPII (Stratagene)

10 pBluescriptII KS- (Stratagene)

pMOSBlue T-vector (Amersham, Buckinghamshire, U.K.)

pCR2.1-TOPO (Invitrogen)

Media

P. rhodozyma strain was maintained routinely in YPD medium (DIFCO, Detroit, U.S.A.).

15 *E. coli* strain was maintained in LB medium (10 g Bacto-trypton, 5 g yeast extract (DIFCO) and 5 g NaCl per liter). NZY medium (5 g NaCl, 2 g MgSO₄·7H₂O, 5 g yeast extract (DIFCO), 10 g NZ amine type A (WAKO, Osaka, Japan) per liter) is used for λ phage propagation in a soft agar (0.7 % agar (WAKO)). When an agar medium was prepared, 1.5 % of agar (WAKO) was supplemented.

20 Methods

Restriction enzymes and T4 DNA ligase were purchased from Takara Shuzo (Ohtsu, Japan).

Isolation of a chromosomal DNA from *P. rhodozyma* was performed by using QIAGEN Genomic Kit (QIAGEN, Hilden, Germany) following the protocol supplied by the manufacturer. Mini-prep of plasmid DNA from transformed *E. coli* was performed with the
 25 Automatic DNA isolation system (PI-50, Kurabo, Co. Ltd., Osaka, Japan). Midi-prep of plasmid DNA from an *E. coli* transformant was performed by using QIAGEN column (QIAGEN). Isolation of λ DNA was performed by Wizard lambda preps DNA purification system (Promega, Madison, U.S.A.) following the protocol prepared by the
 30 manufacturer. A DNA fragment was isolated and purified from agarose by using QIAquick or QIAEX II (QIAGEN). Manipulation of λ phage derivatives was followed by the protocol prepared by the manufacturer (Stratagene).

To clone a partial ACC gene from *P. rhodozyma*, a degenerate PCR method was exploited. Species and accession number to database whose sequence for acetyl-CoA carboxylase were used for multiple alignment analysis are as follows.

	<i>Arabidopsis thaliana</i>	D34630 (DDBJ)
5	<i>Emmericella nidulans</i>	Y15996 (EMBL)
	<i>Gallus gallus</i>	P11029 (Swiss-Prot)
	<i>Glycine max</i>	L48995 (GenBank)
	<i>Homo sapiens</i>	S41121 (PIR)
	<i>Medicago sativa</i>	L25042 (GenBank)
10	<i>Ovis aries</i>	Q28559 (Swiss-Prot)
	<i>Rattus norvegicus</i>	P11497 (Swiss-Prot)
	<i>Saccharomyces cerevisiae</i>	Q00955 (Swiss-Prot)
	<i>Schizosaccharomyces pombe</i>	P78820 (Swiss-Prot)
	<i>Ustilago maydis</i>	S49991 (PIR)

- 15 Two mixed primers whose nucleotide sequences were designed and synthesized based on the common sequence of known acetyl-CoA carboxylase genes from other species: acc9 (sense primer) (SEQ ID NO:4) and acc13 (antisense primer) (SEQ ID NO:5) (in the sequences "n" means nucleotides a, c, g or t; "h" means nucleotides a, c or t, "m" means nucleotides a or c, "k" means nucleotides g or t, and "y" means nucleotides c or t).
- 20 After the PCR reaction of 25 cycles of 95°C for 30 seconds, 45°C for 30 seconds and 72°C for 15 seconds by using ExTaq (Takara Shuzo) as a DNA polymerase and cDNA pool obtained in Example 1 as a template, reaction mixture was applied to agarose gel electrophoresis. One PCR band that had a desired length (0.8 kb) was recovered from the agarose gel and purified by QIAquick (QIAGEN) according to the method by the manufacturer
- 25 and then ligated to pMOSBlue-T-vector (Amersham). After transformation of competent *E. coli* DH5 α , 6 white colonies were selected and plasmids were isolated with Automatic DNA isolation system. As a result of sequencing, it was found that 3 clones had a sequence whose deduced amino acid sequence was similar to known acetyl-CoA carboxylase genes. These isolated cDNA clones were designated as pACC1014 and used for further screening
- 30 study.

Example 3: Isolation of genomic DNA from *P. rhodozyma*

To isolate a genomic DNA from *P. rhodozyma*, QIAGEN genomic kit was used according to the method specified by the manufacturer.

At first, cells of *P. rhodozyma* ATCC96594 strain from 100 ml of overnight culture in YPD medium were harvested by centrifugation (1500 x g for 10 min.) and washed once with TE buffer (10 mM Tris / HCl (pH 8.0) containing 1 mM EDTA). After suspending in 8 ml of Y1 buffer of the QIAGEN genomic kit, lyticase (SIGMA, St. Louis, U.S.A.) was added at the concentration of 2 mg/ml to disrupt cells by enzymatic degradation and the reaction mixture was incubated for 90 min at 30°C and then proceeded to the next extraction step. Finally, 20 µg of genomic DNA was obtained.

Example 4: Southern blot hybridization by using pACC1014 as a probe

Southern blot hybridization was performed to clone a genomic fragment which contains ACC gene from *P. rhodozyma*. Two µg of genomic DNA was digested by *Eco*RI and subjected to agarose gel electrophoresis followed by acidic and alkaline treatment. The denatured DNA was transferred to nylon membrane (Hybond N+, Amersham) by using transblot (Joto Rika, Tokyo, Japan) for an hour. The DNA which was transferred to nylon membrane was fixed by a heat treatment (80°C, 90 min). A probe was prepared by labeling a template DNA (*Eco*RI and *Sal*I -digested pACC1014) with DIG multipriming method (Boehringer Mannheim). Hybridization was performed with the method specified by the manufacturer. As a result, a hybridized band was visualized in the range from 2.0 to 2.3 kilobases (kb).

Example 5: Cloning of a genomic fragment containing the ACC gene

4 µg of the genomic DNA were digested by *Eco*RI and subjected to agarose gel electrophoresis. Then, DNAs with a length within the range from 1.5 to 2.7 kb was recovered by QIAEX II gel extraction kit (QIAGEN) according to the method specified by the manufacturer. The purified DNA was ligated to 0.5 µg of *Eco*RI-digested and CIAP (calf intestine alkaline phosphatase)-treated λZAP II (Stratagene) at 16°C overnight, and packaged by Gigapack III gold packaging extract (Stratagene). The packaged extract was infected to *E. coli* MRF' strain and over-laid with NZY medium poured onto LB agar medium. About 5000 plaques were screened by using *Eco*RI and *Sal*I-digested pACC1014 as a probe. Five plaques were hybridized to the labeled probe.

The *in vivo* excision protocol was applied to these λZAP II derivatives containing putative ACC gene from *P. rhodozyma* by following the instruction manual (Stratagene) to clone the insert fragment into *E. coli* cloning vector, pBluescript SK. Each clone recovered from five positive plaques was subjected for sequencing analysis and it was found that the three of them had the identical sequence to the insert fragment of pACC1014. One of the clone

revealed that this clone contained 5' fragment of *ACC* gene as a result of BLAST X analysis. This clone was named as pACCPvu126 and used for further study.

Example 10: Southern blot hybridization by using pACCPvu126 as a probe

Southern blot hybridization was performed to clone a genomic fragment which covered 5' end of *ACC* gene from *P. rhodozyma*. In a similar manner as Example 7, Southern blot hybridization was performed. A probe was prepared by labeling a template DNA (*Eco*RI-digested pACCPvu116) with DIG multipriming method (Boehringer Mannheim). Hybridization was performed with the method specified by the manufacturer. As a result, a hybridized band whose size was close to 5.0 kb was visualized.

Example 11: Cloning of the genomic clone covering 5' end of *ACC* gene

In a similar manner to Example 8, the genomic fragment containing the insert fragment in pACCPvu126 was cloned by plaque hybridization. The genomic library covering 2.7 to 6.0 kb in length prepared in Example 8 was also used. Twelve positive plaques which hybridized to the insert fragment of pACCPvu126 labeled with DIG were isolated and subjected to in vivo excision to obtain plasmid DNA. As a result of sequencing for thus isolated plasmids, most of the plasmids had the identical sequence to the insert fragment of pACCPvu126. One of the clones was named as pACC204 and used for further study.

Example 12: Cloning of the gapped region between pACC204 and pACC127-17-0.9

As a result of BLAST X analysis against known acetyl-CoA carboxylase genes succeeding to the sequencing study of 3' end of the insert fragment in pACC204 and 5' end of the insert fragment in pACC127-17-0.9, it was suggested that an approximately 0.3 kb fragment could be still missing for a coverage of the entire *ACC* gene. The following PCR primers were synthesized based on the internal sequence of pACC204 and pACC127-17-0.9: acc43 (sense primer) (SEQ ID NO:9) and acc44 (antisense primer) (SEQ ID NO:10).

After the PCR reaction of 25 cycles of 94°C for 15 seconds, 55°C for 30 seconds and 72°C for 15 seconds by using HF polymerase (Clontech) as a DNA polymerase and a genomic DNA obtained in Example 3 as a template, the reaction mixture was applied to agarose gel electrophoresis. One PCR band that had a desired length (0.3 kb) was recovered from the agarose gel and purified by QIAquick (QIAGEN) according to the method by the manufacturer and then cloned into pCR2.1-TOPO (Invitrogen). After transformation of competent *E. coli* TOP10, 6 white colonies were selected and plasmids were isolated with Automatic DNA isolation system. As a result of sequencing, it was found that 5 clones had an

DNA was digested by *Eco*RI and subjected to agarose gel electrophoresis. Then, DNAs with a length within the following range were recovered by QIAEX II gel extraction kit (QIAGEN) according to the method specified by the manufacturer: (1) from 2.7 to 5.0 kb; (2) from 1.4 to 2.7 kb; and (3) from 0.5 to 1.4 kb.

- 5 Each purified DNA was ligated to 0.5 µg of *Eco*RI-digested and CIAP (calf intestine alkaline phosphatase)-treated λZAP II (Stratagene) at 16 °C overnight, and packaged by Giga-pack III gold packaging extract (Stratagene). The packaged extract was infected to *E. coli* MRF' strain and over-laid with NZY medium poured onto LB agar medium. About 5000 plaques were screened by using *Eco*RI-digested pACCStu107 and pACCPvd107 as probes.
- 10 The following candidates were isolated after plaque hybridization study.

- 1) 3 plaques from the 2.7 to 6.0 kb library by using the insert of pACCPvd107 as a probe.
- 2) 3 plaques from the 1.4 to 2.7 kb library by using the insert of pACCStu107 as a probe.
- 3) 21 plaques from the 0.5 to 1.4 kb library by using the insert of pACCStu107 as a probe.

- The *in vivo* excision protocol was applied to these λZAP II derivatives containing putative
- 15 ACC gene from *P. rhodozyma* by following the instruction manual (Stratagene) to clone the insert fragment into *E. coli* cloning vector, pBluescript SK. Each clone recovered from the positive plaques was subjected for sequencing analysis. At least each clone had the putative ACC gene from BLAST X analysis (<http://www.blast.genome.ad.jp/>). The following clones were selected and used for further analysis:

- 20 pACC119-18 having a 6 kb insert and covering the 3' end of the ACC gene;
- pACC119-17-0.6 having a 0.6 kb insert flanking the 5' end of the pACC1224 insert fragment;
- pACC119-17-2 having a 2 kb insert flanking the 5' end of the pACC119-17-0.6 insert fragment; and
- 25 pACC127-17-0.9 having a 0.9kb insert flanking the 5' end of the pACC119-17-2 insert fragment.

As a result of whole sequencing of the entire region of insert fragment in pACC119-18, pACC119-17-0.6, pACC119-17-2 and pACC127-17-0.9, it was suggested that these clones did not cover the 5' end of the ACC gene.

- 30 **Example 9: Cloning of the flanking region of the insert fragment in pACC127-17-0.9 from the genome of *P. rhodozyma* by genome walking method**

PCR primer acc26 (SEQ ID NO:8) was synthesized based on the internal sequence of pACC127-17-0.9 and used for genome walking method.

- In the PCR reaction using acc26 primer, a 2.6 kb PCR band emerged from the genomic
- 35 *Pvu*II library. This PCR band was cloned into pCR2.1-TOPO (Invitrogen) and it was

was named as pACC1224 and used for further study. As a result of whole sequencing of the entire region of insert fragment in pACC1224, it was suggested that this clone contained neither its 5'- nor 3'-end of the ACC gene.

5 **Example 6: Cloning of the flanking region of the insert fragment in pACC1224 from the genome of *P. rhodozyma* by genome walking method**

Two PCR primers were synthesized based on the internal sequence of pACC1224 and used for the genome walking method: acc17 (SEQ ID NO:6) and acc18 (SEQ ID NO:7). The protocol of the instruction manual provided from the supplier (Clontech) was followed for the genome walking method. In the PCR reaction using acc17 primer, a 2.8
10 kb PCR band emerged from the genomic *Stu*I library. In the case of acc18 primer, a 2.2 kb PCR band was produced in the genomic *Pvu*II library. These PCR bands were cloned into pCR2.1-TOPO (Invitrogen) and it was revealed that 2.8 kb PCR band contained a 5' fragment of ACC gene and 2.2 kb PCR band contained 3' fragment of ACC gene, respectively. The clones containing 2.8 kb and 2.2 kb PCR fragment were named as pACC*Stu*107 and
15 pACCPvd107, respectively and used for further study.

Example 7: Southern blot hybridization by using pACC*Stu*107 and pACCPvd107 as probes

Southern blot hybridization was performed to clone a genomic fragment which covered the ACC gene from *P. rhodozyma*. 2 µg of genomic DNA was digested by *Eco*RI and sub-
20 jected to agarose gel electrophoresis followed by acidic and alkaline treatment. The denatured DNA was transferred to nylon membrane (Hybond N+, Amersham) by using transblot (Joto Rika, Tokyo, Japan) for an hour. The DNA which was transferred to nylon membrane was fixed by a heat treatment (80°C, 90 min). A probe was prepared by labeling a template DNA (*Eco*RI -digested pACC*Stu*107 and pACCPvd107) with the DIG multi-
25 priming method (Boehringer Mannheim). Hybridization was performed with the method specified by the manufacturer. As a result, several hybridized bands whose size was close to 2.0 kb, 0.9 kb and 0.6 kb were visualized when the insert fragment in pACC*Stu*107 was used as a probe. In the case that the insert fragment in pACCPvd107 was used as a probe, a hybridized band was visualized in the range from 6.0 kb to 6.5 kb.

30 **Example 8: Cloning of the genomic clone covering the ACC gene**

In a similar manner to Example 5, the genomic fragment containing the insert fragment in pACC*Stu*107 and pACCPvd107 was cloned by plaque hybridization. 4 µg of the genomic

Then, 3.1 kb of the *SacI* fragment containing ribosomal DNA (rDNA) locus (Wery et al., Gene, 184, 89-97, 1997) is inserted downstream of the G418 cassette on thus prepared plasmid. The rDNA fragment exists in multicopies on the chromosome of eukaryote. The integration event via the rDNA fragment would result in multicopied integration onto the chromosome of the host used and this enables the overexpression of foreign genes which are harbored in expression vector.

Subsequently, ACC promoter is inserted in the upstream of ACC terminator to construct of expression vector which functions in *P. rhodozyma*.

Finally, the antisense ACC construct is completed by inserting the 1.5kb of *SfiI* fragment containing antisense ACC into thus prepared expression vector functioning in *P. rhodo-*
10 *zyma*. A similar plasmid construction is disclosed in EP 1,158,051.

Example 15: Transformation of *P. rhodozyma* with an ACC-antisense vector

The ACC-antisense vector thus prepared is transformed into *P. rhodozyma* wild type strain, ATCC96594. The protocol for the biolistic transformation is disclosed in EP 1,158,051.

15 **Example 16: Characterization of antisense ACC recombinant of *P. rhodozyma***

Antisense ACC recombinant of *P. rhodozyma*, ATCC96594 is cultured in 50 ml of YPD medium in 500 ml Erlenmeyer flask at 20°C for 3 days by using their seed culture which grows in 10 ml of YPD medium in test-tubes (21 mm in diameter) at 20°C for 3 days. For analysis of carotenoid produced appropriate volume of culture broth is withdrawn and
20 used for analysis of their growth, productivity of carotenoids, especially astaxanthin. For analysis of growth, optical density at 660 nm is measured by using a UV-1200 photometer (Shimadzu Corp., Kyoto, Japan) in addition to the determination of their dried cell mass by drying up the cells derived from 1 ml of broth after microcentrifugation at 100°C for one day. For the analysis of the content of astaxanthin and total carotenoids, cells are har-
25 vested from 1.0 ml of broth after microcentrifugation and used for the extraction of the carotenoids from cells of *P. rhodozyma* by disruption with glass beads. After extraction, disrupted cells are removed by centrifugation and the resultant is analyzed for carotenoid content with HPLC. The HPLC condition used is as follows: HPLC column: Chrompack Lichrosorb si-60 (4.6 mm, 250 mm), Temperature: room temperature, Eluent: acetone /
30 hexane (18/82) add 1 ml/L of water to eluent, Injection volume: 10 µl, Flow rate: 2.0 ml/min, Detection: UV at 450 nm. A reference sample of astaxanthin can be obtained from Hoffmann La-Roche (Basel, Switzerland).

identical sequence from each other. One of the isolated clones was designated as pACC210.

Example 13: Sequencing of a complete genomic fragment containing ACC gene

pACC204, pACC210, pACC127-17-0.9, pACC119-17-2, pACC119-17-0.6, pACC1224 and
5 pACC119-18 were sequenced with primer walking procedure by using AutoRead sequencing kit (Pharmacia).

As a result of sequencing, the nucleotide sequence comprising 10561 base pairs of the genomic fragment containing the ACC gene from *P. rhodozyma* containing its promoter (1445 base pairs) and terminator (1030 base pairs) was determined (SEQ ID NO:1).

10 The coding region was 8086 base pairs long and consisted of 19 exons and 18 introns. Introns were dispersed all through the coding region without 5' or 3' bias. It was found that an open reading frame (SEQ ID NO:2) consists of 2187 amino acids (SEQ ID NO:3) whose sequence is strikingly similar to the known amino acid sequence of acetyl-CoA carboxylase from other species (56.28% identity to acetyl-CoA carboxylase from *Bmericella nidulans*)
15 as a result of homology search by GENETYX-SV/RC software (Software Development Co., Ltd., Tokyo, Japan).

Fig. 1 depicts a cloned DNA fragment covering ACC gene region on the chromosome of *P. rhodozyma*.

Example 14: Construction of antisense plasmid for ACC gene

20 An antisense gene fragment which covers the entire structure gene for ACC gene is amplified by PCR and then cloned into an integration vector in which the antisense ACC gene is transcribed by its own ACC promoter in *P. rhodozyma*.

The primers include an asymmetrical recognition sequence for the restriction enzyme, *Sfi*I (GGCCNNNNNGGCC) but their asymmetrical hang-over sequence is designed to be
25 different. This enables a directional cloning into expression vector which has the same asymmetrical sequence at their ligation sequence. The use of such a construction is disclosed in EP 1,158,051.

For the promoter and terminator fragment which can drive the transcription of the antisense ACC gene, the ACC promoter and terminator is cloned from the chromosome by
30 using the sequence information listed in SEQ ID NO:1. The ACC terminator fragment is fused to a G418 resistant cassette by ligating the DNA fragment containing the ACC terminator to a G418 resistant cassette of pG418Sa330 (EP 1,035,206) to an appropriate vector such as pBluescriptII KS- (Stratagene).

Claims

1. An isolated polynucleotide comprising a nucleic acid molecule one or more selected from the group consisting of:
- (a) nucleic acid molecules encoding at least the mature form of the polypeptide depicted in SEQ ID NO:3;
 - (b) nucleic acid molecules comprising the coding sequence as depicted in SEQ ID NO:2;
 - (c) nucleic acid molecules whose nucleotide sequence is degenerate as a result of the genetic code to a nucleotide sequence of (a) or (b);
 - (d) nucleic acid molecules encoding a polypeptide derived from the polypeptide encoded by a polynucleotide of (a) to (c) by way of substitution, deletion and/or addition of one or several amino acids of the amino acid sequence of the polypeptide encoded by a nucleotide of (a) to (c);
 - (e) nucleic acid molecules encoding a polypeptide derived from the polypeptide whose sequence has an identity of 56.3 % or more to the amino acid sequence of the polypeptide encoded by a nucleic acid molecule of (a) or (b);
 - (f) nucleic acid molecules comprising a fragment encoded by a nucleic acid molecule of any one of (a) to (e) and having acetyl-CoA carboxylase activity;
 - (g) nucleic acid molecules comprising a polynucleotide having a sequence of a nucleic acid molecule amplified from a *Phaffia* nucleic acid library using the primers depicted in SEQ ID NO:4, 5, and 6;
 - (h) nucleic acid molecules encoding a polypeptide having acetyl-CoA carboxylase activity, wherein said polypeptide is a fragment of a polypeptide encoded by any one of (a) to (g);
 - (i) nucleic acid molecules comprising at least 15 nucleotides of a polynucleotide of any one of (a) to (d);
 - (j) nucleic acid molecules encoding a polypeptide having acetyl-CoA carboxylase activity, wherein said polypeptide is recognized by antibodies that have been raised against a polypeptide encoded by a nucleic acid molecule of any one of (a) to (h);
 - (k) nucleic acid molecules obtainable by screening an appropriate library under stringent conditions with a probe having the sequence of the nucleic acid molecule of any one of (a) to (j), and encoding a polypeptide having acetyl-CoA carboxylase activity;
 - (l) nucleic acid molecules whose complementary strand hybridizes under stringent conditions with a nucleic acid molecule of any one of (a) to (k), and encoding a polypeptide having acetyl-CoA carboxylase activity.
2. An isolated polynucleotide comprising a nucleic acid molecule one or more selected from the group consisting of:

- (m) nucleic acid molecules comprising the nucleotide sequence as depicted in SEQ ID NO:1;
- (n) nucleic acid molecules whose nucleotide sequence is degenerate as a result of the genetic code to a nucleotide sequence of (m);
5. (o) nucleic acid molecules encoding a polypeptide derived from the polypeptide encoded by a polynucleotide of (m) or (n) by way of substitution, deletion and/or addition of one or several amino acids of the amino acid sequence of the polypeptide encoded by a nucleotide of (m) or (n);
- (p) nucleic acid molecules encoding a polypeptide derived from the polypeptide whose
10 sequence has an identity of 56.3 % or more to the amino acid sequence of the polypeptide encoded by a nucleic acid molecule of (m);
- (q) nucleic acid molecules comprising a fragment encoded by a nucleic acid molecule of any one of (m) to (p) and having acetyl-CoA carboxylase activity;
- (r) nucleic acid molecules comprising a polynucleotide having a sequence of a nucleic acid
15 molecule amplified from a *Phaffia* nucleic acid library using the primers depicted in SEQ ID NO:4, 5, and 6;
- (s) nucleic acid molecules encoding a polypeptide having acetyl-CoA carboxylase activity, wherein said polypeptide is a fragment of a polypeptide encoded by any one of (m) to (r);
- (t) nucleic acid molecules comprising at least 15 nucleotides of a polynucleotide of any one
20 of (m) to (o);
- (u) nucleic acid molecules encoding a polypeptide having acetyl-CoA carboxylase activity, wherein said polypeptide is recognized by antibodies that have been raised against a polypeptide encoded by a nucleic acid molecule of any one of (m) to (s);
- (v) nucleic acid molecules obtainable by screening an appropriate library under stringent
25 conditions with a probe having the sequence of the nucleic acid molecule of any one of (m) to (u), and encoding a polypeptide having acetyl-CoA carboxylase activity;
- (w) nucleic acid molecules whose complementary strand hybridizes under stringent conditions with a nucleic acid molecule of any one of (m) to (v), and encoding a polypeptide having acetyl-CoA carboxylase activity.
- 30 3. The isolated polynucleotide of claim 1 or 2, wherein said polynucleotide encodes amino acid sequence which is identified by SEQ ID NO: 3 or has identity of 56.3 % or more with SEQ ID NO: 3.
4. The isolated polynucleotide of any one of claims 1 to 3, wherein said polynucleotide is derived from a strain of *P. rhodozyma* or *Xanthophylomyces dendrorhous*.

5. A method for making a recombinant vector comprising inserting the polynucleotide of any one of claims 1 to 4 into a vector.
6. A recombinant vector containing the polynucleotide of any one of claims 1 to 4 or produced by the method of claim 5.
- 5 7. The vector of claim 6 in which the polynucleotide of any one of claims 1 to 4 is operatively linked to expression control sequences allowing expression in prokaryotic or eukaryotic cells.
8. A method of making a recombinant organism comprising introducing the vector of claim 6 or 7 into a host organism.
- 10 9. The method of claim 8, wherein said host organism is selected from *E. coli*, baculovirus, or *S. cerevisiae*.
10. The recombinant organism containing the vector of claim 6 or 7, or produced by the method of claim 8 or 9.
11. A process for producing a polypeptide having acetyl-CoA carboxylase activity
15 comprising culturing the recombinant organism of claim 10 and recovering the polypeptide from the culture of said recombinant organism.
12. A polypeptide obtainable by the process of claim 11.
13. An antibody that binds specifically to the polypeptide of claim 12.
14. An antisense polynucleotide against the polynucleotide of any one of claims 1 to 4.
- 20 15. A method for making a recombinant vector comprising inserting the polynucleotide of claim 14 into a vector.
16. A recombinant vector containing the polynucleotide of claim 14 or produced by the method of claim 15.
17. The vector of claim 16 in which the polynucleotide of claim 14 is operatively linked to
25 expression control sequences allowing expression in prokaryotic or eukaryotic cells.
18. A method of making a recombinant organism comprising introducing the vector of claim 16 or 17 into a host organism.

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19. The method of claim 18, wherein said host organism is belongs to a strain of *Phaffia rhodozyma* or *Xanthophylomyces dendrorhous*.
20. The recombinant organism containing the vector of claim 16 or 17, or produced by the method of claim 18 or 19.
- 5 21. The recombinant organism of claim 20, wherein said organism is characterized in that whose gene expression of acetyl-CoA carboxylase is reduced compared to the host organism, thereby is capable of producing carotenoids in an enhanced level relative to a host organism.
- 10 22. The recombinant organism according to claim 21, wherein the gene expression of acetyl-CoA carboxylase is reduced by means of the technique selected from antisense technology, site-directed mutagenesis, error prone PCR, or chemical mutagenesis.
23. A process for producing carotenoids, which comprises cultivating the recombinant organism of claim 21.
- 15 24. The process of claim 23, wherein said carotenoids are selected one or more from astaxanthin, β -carotene, lycopene, zeaxanthin, canthaxanthin.
25. The process according to claim 23, wherein the gene expression of acetyl-CoA carboxylase is reduced in the recombinant organism of claim 21 by means of the technique selected from antisense technology, site-directed mutagenesis, error prone PCR, or chemical mutagenesis.
- 20 26. A process for the production of a carotenoid by culturing a microorganism under suitable conditions and, optionally, recovering the resulting carotenoid, wherein the microorganism is characterized in that its gene expression of acetyl-CoA carboxylase is reduced, e.g. by means of the technique selected from antisense technology, site-directed mutagenesis, error prone PCR, or chemical mutagenesis.

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	Asn Val	Leu Ser Phe Phe Pro	Ala Tyr His His Gln	Asp Phe Thr	
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	Ala Ser Glu Val Pro Gly Ser Pro Ile Phe Ile Met Ala Leu Ala Gly			
25	260	265	270	
	tct gct cga cat ttg gag gtc cag ctc ctt gct gat cag tac gga aac			864
	Ser Ala Arg His Leu Glu Val Gln Leu Leu Ala Asp Gln Tyr Gly Asn			
	275	280	285	
30				
	gct atc tct ttg ttc ggt cga gat tgc tct gtt cag cga cga cat cag			912
	Ala Ile Ser Leu Phe Gly Arg Asp Cys Ser Val Gln Arg Arg His Gln			
	290	295	300	
	aag atc att gag gag gct ccc gtc acg atc gct cgt cca gag aga ttc			960
35	Lys Ile Ile Glu Glu Ala Pro Val Thr Ile Ala Arg Pro Glu Arg Phe			
	305	310	315	320
	gaa gag atg gag aag gct gct gtc agg ttg gcc aag tta gta gga tat			1008
	Glu Glu Met Glu Lys Ala Ala Val Arg Leu Ala Lys Leu Val Gly Tyr			
40	325	330	335	
	gtt agt gcc ggt acc gtc gaa tac ctc tac tct cac gcc gac gac tca			1056
	Val Ser Ala Gly Thr Val Glu Tyr Leu Tyr Ser His Ala Asp Asp Ser			
	340	345	350	

	gat ttc gag cag aac gcc att acc acc gct tgg ttg gat ggg ttg atc	1632
	Asp Phe Glu Gln Asn Ala Ile Thr Thr Ala Trp Leu Asp Gly Leu Ile	
	530 535 540	
5	act aac aag ctt aca tct gag agg cct gat cca tca ctg gcc gtt att	1680
	Thr Asn Lys Leu Thr Ser Glu Arg Pro Asp Pro Ser Leu Ala Val Ile	
	545 550 555 560	
	tgt ggt gca att gtg aaa gct cac gtg gct tct gag aac tgt tgg gcc	1728
10	Cys Gly Ala Ile Val Lys Ala His Val Ala Ser Glu Asn Cys Trp Ala	
	565 570 575	
	gaa tac cga cga gta ttg gac aag gga cag gtt ccc tcc aag gac act	1776
	Glu Tyr Arg Arg Val Leu Asp Lys Gly Gln Val Pro Ser Lys Asp Thr	
15	580 585 590	
	ctc aag aca gtg ttc act ctt gat ttc atc tat gag ggt gtt cgg tac	1824
	Leu Lys Thr Val Phe Thr Leu Asp Phe Ile Tyr Glu Gly Val Arg Tyr	
	595 600 605	
20	aat ttc acc gct gct cga gcc tcc ctc aac act tac cga ttg tat cta	1872
	Asn Phe Thr Ala Ala Arg Ala Ser Leu Asn Thr Tyr Arg Leu Tyr Leu	
	610 615 620	
25	aac gga gga aag acc gtg gtg tcc atc cga cct ttg gcc gat ggt gga	1920
	Asn Gly Gly Lys Thr Val Val Ser Ile Arg Pro Leu Ala Asp Gly Gly	
	625 630 635 640	
	atg ctc gtt ctt ctc gat ggc cga tcc cac act ctc tac tgg agg gag	1968
30	Met Leu Val Leu Leu Asp Gly Arg Ser His Thr Leu Tyr Trp Arg Glu	
	645 650 655	
	gaa gtc ggt acc ctc cga att cag gta gac gca aag act tgc ctg att	2016
	Glu Val Gly Thr Leu Arg Ile Gln Val Asp Ala Lys Thr Cys Leu Ile	
	660 665 670	
35	gag cag gag aac gac ccc act cag ctc cga tca ccc tgg cct gga aag	2064
	Glu Gln Glu Asn Asp Pro Thr Gln Leu Arg Ser Pro Ser Pro Gly Lys	
	675 680 685	
40	atc atc cgg ttt ttg gtc gaa agc gga gat cac atc tcc tcc gga gat	2112
	Ile Ile Arg Phe Leu Val Glu Ser Gly Asp His Ile Ser Ser Gly Asp	
	690 695 700	

	ttc ttc ttc ctc gaa ctc aac cct cga ctt caa gtc gag cac cct act	1104
	Phe Phe Phe Leu Glu Leu Asn Pro Arg Leu Gln Val Glu His Pro Thr	
	355 360 365	
5		
	acc gag atg gtc tcg ggt gtc aac ctt ccc gct gct cag ctt cag att	1152
	Thr Glu Met Val Ser Gly Val Asn Leu Pro Ala Ala Gln Leu Gln Ile	
	370 375 380	
10		
	gct atg ggt atc cct ctt tct cga att cgg gat att cga gtc ctc tac	1200
	Ala Met Gly Ile Pro Leu Ser Arg Ile Arg Asp Ile Arg Val Leu Tyr	
	385 390 395 400	
	ggt ctc gat ccc cac act gtt tcc gag atc gac ttc gac agc agc aga	1248
15	Gly Leu Asp Pro His Thr Val Ser Glu Ile Asp Phe Asp Ser Ser Arg	
	405 410 415	
	gcg gag tct gtc cag act cag agg aag cct agg ccc aag ggt cac gtc	1296
	Ala Glu Ser Val Gln Thr Gln Arg Lys Pro Arg Pro Lys Gly His Val	
20	420 425 430	
	att gcc tgt cga atc acg agt gaa aac ccc gat gag ggg ttc aag ccg	1344
	Ile Ala Cys Arg Ile Thr Ser Glu Asn Pro Asp Glu Gly Phe Lys Pro	
	435 440 445	
25		
	tct gcc gga gat atc caa gag ttg aac ttc aga agt aat act aac gtc	1392
	Ser Ala Gly Asp Ile Gln Glu Leu Asn Phe Arg Ser Asn Thr Asn Val	
	450 455 460	
30		
	tgg gga tac ttc tct gtt gga gct act gga gga att cat agt ttc gcc	1440
	Trp Gly Tyr Phe Ser Val Gly Ala Thr Gly Gly Ile His Ser Phe Ala	
	465 470 475 480	
	gat tct caa ttc ggt cac gtg ttt gct tat ggc tcc gac cga acg act	1488
	Asp Ser Gln Phe Gly His Val Phe Ala Tyr Gly Ser Asp Arg Thr Thr	
35	485 490 495	
	gcc aga aag aat atg gtt atc gcc ttg aaa gag ctt tcc att cga gga	1536
	Ala Arg Lys Asn Met Val Ile Ala Leu Lys Glu Leu Ser Ile Arg Gly	
	500 505 510	
40		
	gac ttc cga acc act gtc gag tat ctt atc act ctt ctt gag acg agc	1584
	Asp Phe Arg Thr Thr Val Glu Tyr Leu Ile Thr Leu Leu Glu Thr Ser	
	515 520 525	

	atc tac gct gag gtt gag gtc atg aag atg atc ttg ccc ttg att gcc	2160
	Ile Tyr Ala Glu Val Glu Val Met Lys Met Ile Leu Pro Leu Ile Ala	
	705 710 715 720	
5	cag gag tcc ggt cac gtt cag ttt gtc aag caa gcc ggt gtg acc gtc	2208
	Gln Glu Ser Gly His Val Gln Phe Val Lys Gln Ala Gly Val Thr Val	
	725 730 735	
	gat cct gga gcg att att ggg atc ttg agt ctt gat gac cct acg cga	2256
10	Asp Pro Gly Ala Ile Ile Gly Ile Leu Ser Leu Asp Asp Pro Thr Arg	
	740 745 750	
	gtg aag aag gcg aag ccc ttc gag ggt ctc ctg cct gtg act ggt ctc	2304
	Val Lys Lys Ala Lys Pro Phe Glu Gly Leu Leu Pro Val Thr Gly Leu	
15	755 760 765	
	cct aac ctg ccc ggt aac aga cct cac cag cgg cta cag ttc cag ctt	2352
	Pro Asn Leu Pro Gly Asn Arg Pro His Gln Arg Leu Gln Phe Gln Leu	
	770 775 780	
20		
	gag tcg ata tac tcg gtc ttg gat gga tac gag agt gac tcc act gca	2400
	Glu Ser Ile Tyr Ser Val Leu Asp Gly Tyr Glu Ser Asp Ser Thr Ala	
	785 790 795 800	
25	aca atc ctc cga tca ttc tct gaa aac ctt tat gat cct gat ctt gct	2448
	Thr Ile Leu Arg Ser Phe Ser Glu Asn Leu Tyr Asp Pro Asp Leu Ala	
	805 810 815	
	ttc gga gag gct tta tcc atc att tcc gtc ctt tct ggg aga atg cct	2496
30	Phe Gly Glu Ala Leu Ser Ile Ile Ser Val Leu Ser Gly Arg Met Pro	
	820 825 830	
	gcc gat ctt gag gag agc att cga gag gtc atc agc gaa gct cag tcg	2544
	Ala Asp Leu Glu Glu Ser Ile Arg Glu Val Ile Ser Glu Ala Gln Ser	
	835 840 845	
35		
	aag cct cac gcc gag ttc cct gga tca aag atc ctc aaa gtc gtc gag	2592
	Lys Pro His Ala Glu Phe Pro Gly Ser Lys Ile Leu Lys Val Val Glu	
	850 855 860	
40	cgg tac atc gat aat ttg cga cct cag gag agg gct atg gtc cga act	2640
	Arg Tyr Ile Asp Asn Leu Arg Pro Gln Glu Arg Ala Met Val Arg Thr	
	865 870 875 880	

	cag atc gaa ccc atc gtt ggt att get gag aag aac gtt ggc ggt cct	2688
	Gln Ile Glu Pro Ile Val Gly Ile Ala Glu Lys Asn Val Gly Gly Pro	
	885 890 895	
5	aag ggt tac gcc tct tac gtc tta gct acc atc ctt caa aag ttc ttg	2736
	Lys Gly Tyr Ala Ser Tyr Val Leu Ala Thr Ile Leu Gln Lys Phe Leu	
	900 905 910	
	gcc gtt gag gcc gtt ttt gct act ggt agt gaa gag gcc att gtt ctc	2784
10	Ala Val Glu Ala Val Phe Ala Thr Gly Ser Glu Glu Ala Ile Val Leu	
	915 920 925	
	caa ctt cga gat gaa aac cga gaa tct ttg aac gac gtc ctt ggt ctc	2832
	Gln Leu Arg Asp Glu Asn Arg Glu Ser Leu Asn Asp Val Leu Gly Leu	
15	930 935 940	
	gtc ctg gct cac tcg cgt ctc agc gct cga tcc aag ctt gtt ctc tcc	2880
	Val Leu Ala His Ser Arg Leu Ser Ala Arg Ser Lys Leu Val Leu Ser	
	945 950 955 960	
20	gtc ttt gat ctg atc aag tct atg cag ctc ctc aac aac act gag ggt	2928
	Val Phe Asp Leu Ile Lys Ser Met Gln Leu Leu Asn Asn Thr Glu Gly	
	965 970 975	
25	tct ttc ctt cat aag act atg aaa gcg ctt gcc gac atg ccc acc aag	2976
	Ser Phe Leu His Lys Thr Met Lys Ala Leu Ala Asp Met Pro Thr Lys	
	980 985 990	
	gct cct ttg gcc agc aag gtg tct ttg aag gct cgg gaa att ctt atc	3024
30	Ala Pro Leu Ala Ser Lys Val Ser Leu Lys Ala Arg Glu Ile Leu Ile	
	995 1000 1005	
	tct tgc tct ctt ccc tct tac gag gag agg ttg ttc cag atg gaa	3069
	Ser Cys Ser Leu Pro Ser Tyr Glu Glu Arg Leu Phe Gln Met Glu	
	1010 1015 1020	
35	aag atc ctt aac tct tct gtc acc act tct tac tac gga gag act	3114
	Lys Ile Leu Asn Ser Ser Val Thr Thr Ser Tyr Tyr Gly Glu Thr	
	1025 1030 1035	
40	gga ggt gga cac aga aac cct tcg gtt gat gtt ctg act gag atc	3159
	Gly Gly Gly His Arg Asn Pro Ser Val Asp Val Leu Thr Glu Ile	
	1040 1045 1050	

	tca aac	tct cga ttc acc gtc	tac gat gtc ctg tcc	tcc ttc ttc	3204
	Ser Asn	Ser Arg Phe Thr Val	Tyr Asp Val Leu Ser	Ser Phe Phe	
	1055	1060	1065		
5	aag cac	gat gat cct tgg att	gtt ctt gct agt ttg	acc gtc tac	3249
	Lys His	Asp Asp Pro Trp Ile	Val Leu Ala Ser Leu	Thr Val Tyr	
	1070	1075	1080		
	gtt ctt	cga gct tac cga gag	tac agt att ctt gat	atg caa cat	3294
10	Val Leu	Arg Ala Tyr Arg Glu	Tyr Ser Ile Leu Asp	Met Gln His	
	1085	1090	1095		
	gag caa	ggc cag gat ggc gct	gct gga gtc atc act	tgg cga ttc	3339
	Glu Gln	Gly Gln Asp Gly Ala	Ala Gly Val Ile Thr	Trp Arg Phe	
15	1100	1105	1110		
	aag ctc	aac cag ccc atc gct	gag tct tct act ccc	cga gtt gac	3384
	Lys Leu	Asn Gln Pro Ile Ala	Glu Ser Ser Thr Pro	Arg Val Asp	
	1115	1120	1125		
20	tcg aat	cga gac gtt tac cga	gtc ggt tcg ctt tct	gat ttg acc	3429
	Ser Asn	Arg Asp Val Tyr Arg	Val Gly Ser Leu Ser	Asp Leu Thr	
	1130	1135	1140		
25	tac aag	atc aag cag agt cag	acc gag ccc ctc cga	gct ggt gtc	3474
	Tyr Lys	Ile Lys Gln Ser Gln	Thr Glu Pro Leu Arg	Ala Gly Val	
	1145	1150	1155		
	atg acg	agc ttc aac aac ttg	aag gag gtt cag gac	gga ctc ttg	3519
30	Met Thr	Ser Phe Asn Asn Leu	Lys Glu Val Gln Asp	Gly Leu Leu	
	1160	1165	1170		
	aat gtt	ctg tct ttc ttc cct	gct tac cat cat caa	gat ttc act	3564
	Asn Val	Leu Ser Phe Phe Pro	Ala Tyr His His Gln	Asp Phe Thr	
	1175	1180	1185		
35	caa cga	cat ggt cag gac agt	gcc atg ccc aac gtt	ctc aac att	3609
	Gln Arg	His Gly Gln Asp Ser	Ala Met Pro Asn Val	Leu Asn Ile	
	1190	1195	1200		
40	gct atc	cgg gct ttc gag gag	aag gac gac atg tct	gat ctt gat	3654
	Ala Ile	Arg Ala Phe Glu Glu	Lys Asp Asp Met Ser	Asp Leu Asp	
	1205	1210	1215		

	tcg gcc	aag agt gtt gag tcg	ctg gta atg cag atg	tct gcc gag	3699
	Trp Ala	Lys Ser Val Glu Ser	Leu Val Met Gln Met	Ser Ala Glu	
	1220	1225	1230		
5	atc cag	aag aag gga att cga	cga gtt acc ttc ttg	gtt tgc cga	3744
	Ile Gln	Lys Lys Gly Ile Arg	Arg Val Thr Phe Leu	Val Cys Arg	
	1235	1240	1245		
10	aag ggc	gtt tac ccc tcc tac	ttc acc ttc aga caa	gag ggt gcc	3789
	Lys Gly	Val Tyr Pro Ser Tyr	Phe Thr Phe Arg Gln	Glu Gly Ala	
	1250	1255	1260		
15	cag ggc	ccc tgg aga gag gag	gag aag att cga aac	atc gag cct	3834
	Gln Gly	Pro Trp Arg Glu Glu	Glu Lys Ile Arg Asn	Ile Glu Pro	
	1265	1270	1275		
20	gct cta	gcc agt cag ctt gag	ctc aac cga ctc tcg	aat ttc aag	3879
	Ala Leu	Ala Ser Gln Leu Glu	Leu Asn Arg Leu Ser	Asn Phe Lys	
	1280	1285	1290		
	gtc acc	cct atc ttc gta gac	aac aga cag atc cac	atc tac aag	3924
	Val Thr	Pro Ile Phe Val Asp	Asn Arg Gln Ile His	Ile Tyr Lys	
	1295	1300	1305		
25	gga gtg	ggt aag gag aac tct	tcc gat gtt cga ttc	ttt atc cgg	3969
	Gly Val	Gly Lys Glu Asn Ser	Ser Asp Val Arg Phe	Phe Ile Arg	
	1310	1315	1320		
30	gct ttg	gtt cga cct gga cgg	gtc cag gga tcg atg	aag gct gcc	4014
	Ala Leu	Val Arg Pro Gly Arg	Val Gln Gly Ser Met	Lys Ala Ala	
	1325	1330	1335		
	gag tat	ctc atc tcc gag tgc	gat cga ctg ctc act	gat atc ctg	4059
	Glu Tyr	Leu Ile Ser Glu Cys	Asp Arg Leu Leu Thr	Asp Ile Leu	
	1340	1345	1350		
35	gac gcc	ttg gag gtt gtt gga	gcc gag act cga aac	gcc gat tgc	4104
	Asp Ala	Leu Glu Val Val Gly	Ala Glu Thr Arg Asn	Ala Asp Cys	
	1355	1360	1365		
40	aac cat	gtt gga att aac ttc	atc tat aac gtt ctt	gtc gac ttc	4149
	Asn His	Val Gly Ile Asn Phe	Ile Tyr Asn Val Leu	Val Asp Phe	
	1370	1375	1380		

	gac gac	gtc cag gag gcc ctt	gcc ggg ttc att gag	agg cac gga	4194
	Asp Asp	Val Gln Glu Ala Leu	Ala Gly Phe Ile Glu	Arg His Gly	
	1385	1390	1395		
5	aag agg	ctt tgg cga ctt cga	gtg acc gct tct gaa	atc cga atg	4239
	Lys Arg	Leu Trp Arg Leu Arg	Val Thr Ala Ser Glu	Ile Arg Met	
	1400	1405	1410		
	gtt ctt	gag gac gac gag ggt	aac gtc acc ccc atc	cga tgc tgc	4284
10	Val Leu	Glu Asp Asp Glu Gly	Asn Val Thr Pro Ile	Arg Cys Cys	
	1415	1420	1425		
	att gag	aac gtt tct ggt ttc	gtc gtg aag tac cac	gcc tac cag	4329
	Ile Glu	Asn Val Ser Gly Phe	Val Val Lys Tyr His	Ala Tyr Gln	
15	1430	1435	1440		
	gag gtt	gag acc gag aag ggt	act acc atc ttg aag	tca atc gga	4374
	Glu Val	Glu Thr Glu Lys Gly	Thr Thr Ile Leu Lys	Ser Ile Gly	
	1445	1450	1455		
20					
	gac ctt	gga cct ctt cac ctt	cag cct gtc aac cat	gct tac cag	4419
	Asp Leu	Gly Pro Leu His Leu	Gln Pro Val Asn His	Ala Tyr Gln	
	1460	1465	1470		
	acc aag	aac agt ctt cag ccc	cga cga tac cag gct	cac ttg gtt	4464
25	Thr Lys	Asn Ser Leu Gln Pro	Arg Arg Tyr Gln Ala	His Ile Val	
	1475	1480	1485		
	gga acg	act tac gtc tac gac	tac ccc gat ctc ttc	gtt cag agt	4509
30	Gly Thr	Thr Tyr Val Tyr Asp	Tyr Pro Asp Leu Phe	Val Gln Ser	
	1490	1495	1500		
	ttg cgc	aag gtt tgg gct gag	gct gct gct aag att	cct cac ctc	4554
	Leu Arg	Lys Val Trp Ala Glu	Ala Ala Ala Lys Ile	Pro His Leu	
	1505	1510	1515		
35					
	cgg gtg	cct agc gag cct ctt	acc gct acc gag ttg	gtt ctc gat	4599
	Arg Val	Pro Ser Glu Pro Leu	Thr Ala Thr Glu Leu	Val Leu Asp	
	1520	1525	1530		
	gag aac	aac gag ctt cag gag	gtc gag cga cct ccg	ggc tcc aac	4644
40	Glu Asn	Asn Glu Leu Gln Glu	Val Glu Arg Pro Pro	Gly Ser Asn	
	1535	1540	1545		

	tcg tgt	ggc atg gtc gcc tgg	atc ttc act atg ctc	act ccc gag	4689
	Ser Cys	Gly Met Val Ala Trp	Ile Phe Thr Met Leu	Thr Pro Glu	
	1550	1555	1560		
5	tat ccc	aag ggt cga cga gta	gtt gcc att gcc aac	gat atc acc	4734
	Tyr Pro	Lys Gly Arg Arg Val	Val Ala Ile Ala Asn	Asp Ile Thr	
	1565	1570	1575		
10	ttc aag	att gga tcc ttt ggt	cct aag gaa gac gat	tac ttc ttc	4779
	Phe Lys	Ile Gly Ser Phe Gly	Pro Lys Glu Asp Asp	Tyr Phe Phe	
	1580	1585	1590		
15	aag gct	act gaa att gcc aag	aag ctg ggc ctt cct	cga att tac	4824
	Lys Ala	Thr Glu Ile Ala Lys	Lys Leu Gly Leu Pro	Arg Ile Tyr	
	1595	1600	1605		
20	ctc tct	gcc aac agt gga gct	aga ctc ggt atc gcg	gag gag ctc	4869
	Leu Ser	Ala Asn Ser Gly Ala	Arg Leu Gly Ile Ala	Glu Glu Leu	
	1610	1615	1620		
	ttg cac	atc ttc aag gcg gcc	ttc gtt gac ccc gca	aag cct tcc	4914
	Leu His	Ile Phe Lys Ala Ala	Phe Val Asp Pro Ala	Lys Pro Ser	
	1625	1630	1635		
25	atg ggt	att aag tat cta tac	ttg acc cct gaa act	tta tcc act	4959
	Met Gly	Ile Lys Tyr Leu Tyr	Leu Thr Pro Glu Thr	Leu Ser Thr	
	1640	1645	1650		
30	ctt gcc	aag aag gga tcc agc	gtc acc act gag gag	atc gag gat	5004
	Leu Ala	Lys Lys Gly Ser Ser	Val Thr Thr Glu Glu	Ile Glu Asp	
	1655	1660	1665		
	gac ggc	gag cga cga cac aag	atc acc gcc atc atc	ggt ctt gca	5049
	Asp Gly	Glu Arg Arg His Lys	Ile Thr Ala Ile Ile	Gly Leu Ala	
	1670	1675	1680		
35	gag ggt	ttg gga gtt gag tct	ctt cga gga tcc ggt	ctt att gct	5094
	Glu Gly	Leu Gly Val Glu Ser	Leu Arg Gly Ser Gly	Leu Ile Ala	
	1685	1690	1695		
40	gga gcc	acc act cga gct tac	gag gag gga atc ttc	acc atc tct	5139
	Gly Ala	Thr Thr Arg Ala Tyr	Glu Glu Gly Ile Phe	Thr Ile Ser	
	1700	1705	1710		

	oto gtt	act gcc cga tgg gtc	ggt atc gga gct tac	ttg gtt cga	5184
	Leu Val	Thr Ala Arg Ser Val	Gly Ile Gly Ala Tyr	Leu Val Arg	
	1715	1720	1725		
5	ttg ggt	cag cga gct att cag	ggt gaa ggc aac cct	atg atc ctt	5229
	Leu Gly	Gln Arg Ala Ile Gln	Val Glu Gly Asn Pro	Met Ile Leu	
	1730	1735	1740		
	act gga	gct cag tct ctc aac	aag gtg ctt gga cga	gag gtt tac	5274
10	Thr Gly	Ala Gln Ser Leu Asn	Lys Val Leu Gly Arg	Glu Val Tyr	
	1745	1750	1755		
	act tcc	aac ctt cag ctt gga	gga acc cag att atg	gcc cga aac	5319
15	Thr Ser	Asn Leu Gln Leu Gly	Gly Thr Gln Ile Met	Ala Arg Asn	
	1760	1765	1770		
	ggt acc	acg cat ctc gtc gct	gaa tct gat ctc gat	ggt gct ctc	5364
	Gly Thr	Thr His Leu Val Ala	Glu Ser Asp Leu Asp	Gly Ala Leu	
	1775	1780	1785		
20	aag gtc	atc cag tgg ctc tgg	tat gtg ccc gag cga	aag ggc aag	5409
	Lys Val	Ile Gln Trp Leu Ser	Tyr Val Pro Glu Arg	Lys Gly Lys	
	1790	1795	1800		
25	gcc att	cct atc tgg cct tcc	gag gac cct tgg gac	cga act gtg	5454
	Ala Ile	Pro Ile Trp Pro Ser	Glu Asp Pro Trp Asp	Arg Thr Val	
	1805	1810	1815		
	acc tac	gag cct ccc cga ggt	cct tac gat cct cga	tgg ttg ctt	5499
30	Thr Tyr	Glu Pro Pro Arg Gly	Pro Tyr Asp Pro Arg	Trp Leu Leu	
	1820	1825	1830		
	gaa gga	aag ccg gat gaa ggc	ttg act ggt ctt ttc	gac aag gga	5544
	Glu Gly	Lys Pro Asp Glu Gly	Leu Thr Gly Leu Phe	Asp Lys Gly	
	1835	1840	1845		
35	tct ttc	atg gag acc ctt gga	gat tgg gcc aag act	atc gtc acc	5589
	Ser Phe	Met Glu Thr Leu Gly	Asp Trp Ala Lys Thr	Ile Val Thr	
	1850	1855	1860		
40	ggt cga	gcc cga ctg gga ggc	att cct atg ggt gtt	att gct gtc	5634
	Gly Arg	Ala Arg Leu Gly Gly	Ile Pro Met Gly Val	Ile Ala Val	
	1865	1870	1875		

	gaa acc	agg acg acc	gag aag	atc atc	gct gcc	gat cct	gcc aac	5679
	Glu Thr	Arg Thr Thr	Glu Lys	Ile Ile	Ala Ala	Asp Pro	Ala Asn	
	1880		1885			1890		
5	cct gca	gct ttc	gag caa	aag att	atg gag	gct ggt	cag gtt	5724
	Pro Ala	Ala Phe	Glu Gln	Lys Ile	Met Glu	Ala Gly	Gln Val	Trp
	1895		1900			1905		
	aac ccc	aac gct	gct tac	aag acc	gct caa	tcc atc	ttt gat	5769
10	Asn Pro	Asn Ala	Ala Tyr	Lys Thr	Ala Gln	Ser Ile	Phe Asp	Ile
	1910		1915			1920		
	aac aag	gag ggt	ctt cct	ttg atg	atc ctt	gcc aac	atc cga	5814
15	Asn Lys	Glu Gly	Leu Pro	Leu Met	Ile Leu	Ala Asn	Ile Arg	Gly
	1925		1930			1935		
	ttc tct	gga gga	cag ggt	gat atg	ttt gac	gct atc	ctc aag	5859
	Phe Ser	Gly Gly	Gln Gly	Asp Met	Phe Asp	Ala Ile	Leu Lys	Gln
	1940		1945			1950		
20	ggt tct	aag atc	gtt gac	ggt ctc	tcg aac	ttc aag	cag cca	5904
	Gly Ser	Lys Ile	Val Asp	Gly Leu	Ser Asn	Phe Lys	Gln Pro	Val
	1955		1960			1965		
25	ttc gtc	tat gtt	gtc ccc	aac gga	gag ctt	cgt gga	gga gct	5949
	Phe Val	Tyr Val	Val Pro	Asn Gly	Glu Leu	Arg Gly	Gly Ala	Trp
	1970		1975			1980		
	gtc gtg	ttg gat	cct act	atc aac	ctt gcc	aag atg	gag atg	5994
30	Val Val	Leu Asp	Pro Thr	Ile Asn	Leu Ala	Lys Met	Glu Met	Tyr
	1985		1990			1995		
	gct gat	gaa acc	gct cga	gga gga	att ctc	gag ccg	gaa ggt	6039
	Ala Asp	Glu Thr	Ala Arg	Gly Gly	Ile Leu	Glu Pro	Glu Gly	Ile
	2000		2005			2010		
35	gtt gag	atc aag	ttc cga	cga gac	aag gtc	atc gct	acc atg	6084
	Val Glu	Ile Lys	Phe Arg	Arg Asp	Lys Val	Ile Ala	Thr Met	Glu
	2015		2020			2025		
40	cga ttg	gac gag	acc tat	gcc tct	ctc aaa	gct gcc	tcg aac	6129
	Arg Leu	Asp Glu	Thr Tyr	Ala Ser	Leu Lys	Ala Ala	Ser Asn	Asp
	2030		2035			2040		

	tca acc	aag tct gcg gag gag	cga gct aag agt gct	gag cta ctc	6174
	Ser Thr	Lys Ser Ala Glu Glu	Arg Ala Lys Ser Ala	Glu Leu Leu	
	2045	2050	2055		
5	aag gca	aga gag act cta ctt	caa ccg acg tac ttg	cag att gca	6219
	Lys Ala	Arg Glu Thr Leu Leu	Gln Pro Thr Tyr Leu	Gln Ile Ala	
	2060	2065	2070		
	cac ctt	tac gct gat ctc cat	gat cgt gtc gga cga	atg gag gcc	6264
10	His Leu	Tyr Ala Asp Leu His	Asp Arg Val Gly Arg	Met Glu Ala	
	2075	2080	2085		
	aag ggt	tgc gcg aag cga gct	gtc tgg gct gag gct	cga cga ttc	6309
	Lys Gly	Cys Ala Lys Arg Ala	Val Trp Ala Glu Ala	Arg Arg Phe	
15	2090	2095	2100		
	ttc tac	tgg cga ctt cga cga	cgt ctc aac gat gag	cac atc ctg	6354
	Phe Tyr	Trp Arg Leu Arg Arg	Arg Leu Asn Asp Glu	His Ile Leu	
	2105	2110	2115		
20	tct aag	ttc gct gct gcc aac	ccg gat ctt act ctc	gag gag cga	6399
	Ser Lys	Phe Ala Ala Ala Asn	Pro Asp Leu Thr Leu	Glu Glu Arg	
	2120	2125	2130		
25	caa aac	att ctc gac tct gtc	gtc cag act gac ctc	act gat gac	6444
	Gln Asn	Ile Leu Asp Ser Val	Val Gln Thr Asp Leu	Thr Asp Asp	
	2135	2140	2145		
	cga gcc	acc gct gaa tgg att	gag cag tct gca gaa	gag att gct	6489
30	Arg Ala	Thr Ala Glu Trp Ile	Glu Gln Ser Ala Glu	Glu Ile Ala	
	2150	2155	2160		
	gct gcc	gtt gcc gaa gtc cga	tcc acc tac gtg tcg	aat aag att	6534
	Ala Ala	Val Ala Glu Val Arg	Ser Thr Tyr Val Ser	Asn Lys Ile	
	2165	2170	2175		
35	atc agc	ttc gcc gag acg gag	cga gct gga gcg ttg	cag ggc ttg	6579
	Ile Ser	Phe Ala Glu Thr Glu	Arg Ala Gly Ala Leu	Gln Gly Leu	
	2180	2185	2190		
40	gtc gct	gtc ttg agc act ttg	aat gcg gaa gac aag	aag gcc ctt	6624
	Val Ala	Val Leu Ser Thr Leu	Asn Ala Glu Asp Lys	Lys Ala Leu	
	2195	2200	2205		

gtt tct agc ctc ggt ctc taa
Val Ser Ser Leu Gly Leu
2210

6645

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<212> PRT
<213> Phaffia rhodozyma

10

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1 5 10 15

15

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20 25 30

20 Asp Gly His Thr Val Ile Thr Lys Val Leu Ile Ala Asn Asn Gly Ile
35 40 45

Ala Ala Val Lys Glu Ile Arg Ser Val Arg Lys Trp Ala Tyr Glu Thr
50 55 60

25 Phe Gly Asp Glu Arg Ala Ile Glu Phe Thr Val Met Ala Thr Pro Glu
65 70 75 80

Asp Leu Lys Val Asn Cys Asp Tyr Ile Arg Met Ala Asp Arg Val Val
85 90 95

30

Glu Val Pro Gly Gly Thr Asn Asn Asn Asn His Ser Asn Val Asp Leu
100 105 110

Ile Val Asp Ile Ala Glu Arg Phe Asn Ile His Ala Val Trp Ala Gly
115 120 125

35

Trp Gly His Ala Ser Glu Asn Pro Arg Leu Pro Glu Ser Leu Ala Ala
130 135 140

40 Ser Lys Asn Lys Ile Val Phe Ile Gly Pro Pro Gly Ser Ala Met Arg
145 150 155 160

Ser Leu Gly Asp Lys Ile Ser Ser Thr Ile Val Ala Gln Ser Ala Gln
165 170 175

40

	Gly Leu Asp Pro His Thr Val Ser Glu Ile Asp Phe Asp Ser Ser Arg		
	405	410	415
5	Ala Glu Ser Val Gln Thr Gln Arg Lys Pro Arg Pro Lys Gly His Val		
	420	425	430
	Ile Ala Cys Arg Ile Thr Ser Glu Asn Pro Asp Glu Gly Phe Lys Pro		
	435	440	445
10	Ser Ala Gly Asp Ile Gln Glu Leu Asn Phe Arg Ser Asn Thr Asn Val		
	450	455	460
	Trp Gly Tyr Phe Ser Val Gly Ala Thr Gly Gly Ile His Ser Phe Ala		
	465	470	475 480
15	Asp Ser Gln Phe Gly His Val Phe Ala Tyr Gly Ser Asp Arg Thr Thr		
	485	490	495
	Ala Arg Lys Asn Met Val Ile Ala Leu Lys Glu Leu Ser Ile Arg Gly		
20	500	505	510
	Asp Phe Arg Thr Thr Val Glu Tyr Leu Ile Thr Leu Leu Glu Thr Ser		
	515	520	525
25	Asp Phe Glu Gln Asn Ala Ile Thr Thr Ala Trp Leu Asp Gly Leu Ile		
	530	535	540
	Thr Asn Lys Leu Thr Ser Glu Arg Pro Asp Pro Ser Leu Ala Val Ile		
	545	550	555 560
30			
	Cys Gly Ala Ile Val Lys Ala His Val Ala Ser Glu Asn Cys Trp Ala		
	565	570	575
35	Glu Tyr Arg Arg Val Leu Asp Lys Gly Gln Val Pro Ser Lys Asp Thr		
	580	585	590
	Leu Lys Thr Val Phe Thr Leu Asp Phe Ile Tyr Glu Gly Val Arg Tyr		
	595	600	605
40			
	Asn Phe Thr Ala Ala Arg Ala Ser Leu Asn Thr Tyr Arg Leu Tyr Leu		
	610	615	620

	Asn Gly Gly Lys Thr Val Val Ser Ile Arg Pro Leu Ala Asp Gly Gly	
	625	630 635 640
5	Met Leu Val Leu Leu Asp Gly Arg Ser His Thr Leu Tyr Trp Arg Glu	
	645	650 655
	Glu Val Gly Thr Leu Arg Ile Gln Val Asp Ala Lys Thr Cys Leu Ile	
	660	665 670
10	Glu Gln Glu Asn Asp Pro Thr Gln Leu Arg Ser Pro Ser Pro Gly Lys	
	675	680 685
	Ile Ile Arg Phe Leu Val Glu Ser Gly Asp His Ile Ser Ser Gly Asp	
	690	695 700
15	Ile Tyr Ala Glu Val Glu Val Met Lys Met Ile Leu Pro Leu Ile Ala	
	705	710 715 720
	Gln Glu Ser Gly His Val Gln Phe Val Lys Gln Ala Gly Val Thr Val	
20	725	730 735
	Asp Pro Gly Ala Ile Ile Gly Ile Leu Ser Leu Asp Asp Pro Thr Arg	
	740	745 750
25	Val Lys Lys Ala Lys Pro Phe Glu Gly Leu Leu Pro Val Thr Gly Leu	
	755	760 765
	Pro Asn Leu Pro Gly Asn Arg Pro His Gln Arg Leu Gln Phe Gln Leu	
	770	775 780
30	Glu Ser Ile Tyr Ser Val Leu Asp Gly Tyr Glu Ser Asp Ser Thr Ala	
	785	790 795 800
35	Thr Ile Leu Arg Ser Phe Ser Glu Asn Leu Tyr Asp Pro Asp Leu Ala	
	805	810 815
	Phe Gly Glu Ala Leu Ser Ile Ile Ser Val Leu Ser Gly Arg Met Pro	
	820	825 830
40	Ala Asp Leu Glu Glu Ser Ile Arg Glu Val Ile Ser Glu Ala Gln Ser	
	835	840 845

	Lys Pro His Ala Glu Phe Pro Gly Ser Lys Ile Leu Lys Val Val Glu	
	850	855 860
5	Arg Tyr Ile Asp Asn Leu Arg Pro Gln Glu Arg Ala Met Val Arg Thr	
	865	870 875 880
	Gln Ile Glu Pro Ile Val Gly Ile Ala Glu Lys Asn Val Gly Gly Pro	
	885	890 895
10	Lys Gly Tyr Ala Ser Tyr Val Leu Ala Thr Ile Leu Gln Lys Phe Leu	
	900	905 910
	Ala Val Glu Ala Val Phe Ala Thr Gly Ser Glu Glu Ala Ile Val Leu	
	915	920 925
15	Gln Leu Arg Asp Glu Asn Arg Glu Ser Leu Asn Asp Val Leu Gly Leu	
	930	935 940
	Val Leu Ala His Ser Arg Leu Ser Ala Arg Ser Lys Leu Val Leu Ser	
20	945	950 955 960
	Val Phe Asp Leu Ile Lys Ser Met Gln Leu Leu Asn Asn Thr Glu Gly	
	965	970 975
25	Ser Phe Leu His Lys Thr Met Lys Ala Leu Ala Asp Met Pro Thr Lys	
	980	985 990
	Ala Pro Leu Ala Ser Lys Val Ser Leu Lys Ala Arg Glu Ile Leu Ile	
	995	1000 1005
30	Ser Cys Ser Leu Pro Ser Tyr Glu Glu Arg Leu Phe Gln Met Glu	
	1010	1015 1020
35	Lys Ile Leu Asn Ser Ser Val Thr Thr Ser Tyr Tyr Gly Glu Thr	
	1025	1030 1035
	Gly Gly Gly His Arg Asn Pro Ser Val Asp Val Leu Thr Glu Ile	
	1040	1045 1050
40	Ser Asn Ser Arg Phe Thr Val Tyr Asp Val Leu Ser Ser Phe Phe	
	1055	1060 1065

	Lys	His	Asp	Asp	Pro	Trp	Ile	Val	Leu	Ala	Ser	Leu	Thr	Val	Tyr	
	1070						1075					1080				
5	Val	Leu	Arg	Ala	Tyr	Arg	Glu	Tyr	Ser	Ile	Leu	Asp	Met	Gln	His	
	1085						1090					1095				
	Glu	Gln	Gly	Gln	Asp	Gly	Ala	Ala	Gly	Val	Ile	Thr	Trp	Arg	Phe	
	1100						1105					1110				
10	Lys	Leu	Asn	Gln	Pro	Ile	Ala	Glu	Ser	Ser	Thr	Pro	Arg	Val	Asp	
	1115						1120					1125				
	Ser	Asn	Arg	Asp	Val	Tyr	Arg	Val	Gly	Ser	Leu	Ser	Asp	Leu	Thr	
	1130						1135					1140				
15	Tyr	Lys	Ile	Lys	Gln	Ser	Gln	Thr	Glu	Pro	Leu	Arg	Ala	Gly	Val	
	1145						1150					1155				
	Met	Thr	Ser	Phe	Asn	Asn	Leu	Lys	Glu	Val	Gln	Asp	Gly	Leu	Leu	
20	1160						1165					1170				
	Asn	Val	Leu	Ser	Phe	Phe	Pro	Ala	Tyr	His	His	Gln	Asp	Phe	Thr	
	1175						1180					1185				
25	Gln	Arg	His	Gly	Gln	Asp	Ser	Ala	Met	Pro	Asn	Val	Leu	Asn	Ile	
	1190						1195					1200				
	Ala	Ile	Arg	Ala	Phe	Glu	Glu	Lys	Asp	Asp	Met	Ser	Asp	Leu	Asp	
	1205						1210					1215				
30																
	Trp	Ala	Lys	Ser	Val	Glu	Ser	Leu	Val	Met	Gln	Met	Ser	Ala	Glu	
	1220						1225					1230				
35	Ile	Gln	Lys	Lys	Gly	Ile	Arg	Arg	Val	Thr	Phe	Leu	Val	Cys	Arg	
	1235						1240					1245				
	Lys	Gly	Val	Tyr	Pro	Ser	Tyr	Phe	Thr	Phe	Arg	Gln	Glu	Gly	Ala	
	1250						1255					1260				
40																
	Gln	Gly	Pro	Trp	Arg	Glu	Glu	Glu	Lys	Ile	Arg	Asn	Ile	Glu	Pro	
	1265						1270					1275				

	Ala Leu	Ala Ser Gln Leu Glu	Leu Asn Arg Leu Ser	Asn Phe Lys
	1280		1285	1290
5	Val Thr	Pro Ile Phe Val Asp	Asn Arg Gln Ile His	Ile Tyr Lys
	1295		1300	1305
	Gly Val	Gly Lys Glu Asn Ser	Ser Asp Val Arg Phe	Phe Ile Arg
	1310		1315	1320
10	Ala Leu	Val Arg Pro Gly Arg	Val Gln Gly Ser Met	Lys Ala Ala
	1325		1330	1335
	Glu Tyr	Leu Ile Ser Glu Cys	Asp Arg Leu Leu Thr	Asp Ile Leu
	1340		1345	1350
15	Asp Ala	Leu Glu Val Val Gly	Ala Glu Thr Arg Asn	Ala Asp Cys
	1355		1360	1365
	Asn His	Val Gly Ile Asn Phe	Ile Tyr Asn Val Leu	Val Asp Phe
20	1370		1375	1380
	Asp Asp	Val Gln Glu Ala Leu	Ala Gly Phe Ile Glu	Arg His Gly
	1385		1390	1395
25	Lys Arg	Leu Trp Arg Leu Arg	Val Thr Ala Ser Glu	Ile Arg Met
	1400		1405	1410
	Val Leu	Glu Asp Asp Glu Gly	Asn Val Thr Pro Ile	Arg Cys Cys
	1415		1420	1425
30	Ile Glu	Asn Val Ser Gly Phe	Val Val Lys Tyr His	Ala Tyr Gln
	1430		1435	1440
35	Glu Val	Glu Thr Glu Lys Gly	Thr Thr Ile Leu Lys	Ser Ile Gly
	1445		1450	1455
	Asp Leu	Gly Pro Leu His Leu	Gln Pro Val Asn His	Ala Tyr Gln
	1460		1465	1470
40	Thr Lys	Asn Ser Leu Gln Pro	Arg Arg Tyr Gln Ala	His Leu Val
	1475		1480	1485

	Gly Thr	Thr Tyr Val Tyr Asp	Tyr Pro Asp Leu Phe	Val Gln Ser
	1490	1495	1500	
5	Leu Arg	Lys Val Trp Ala Glu	Ala Ala Ala Lys Ile	Pro His Leu
	1505	1510	1515	
	Arg Val	Pro Ser Glu Pro Leu	Thr Ala Thr Glu Leu	Val Leu Asp
	1520	1525	1530	
10	Glu Asn	Asn Glu Leu Gln Glu	Val Glu Arg Pro Pro	Gly Ser Asn
	1535	1540	1545	
	Ser Cys	Gly Met Val Ala Trp	Ile Phe Thr Met Leu	Thr Pro Glu
	1550	1555	1560	
15	Tyr Pro	Lys Gly Arg Arg Val	Val Ala Ile Ala Asn	Asp Ile Thr
	1565	1570	1575	
	Phe Lys	Ile Gly Ser Phe Gly	Pro Lys Glu Asp Asp	Tyr Phe Phe
20	1580	1585	1590	
	Lys Ala	Thr Glu Ile Ala Lys	Lys Leu Gly Leu Pro	Arg Ile Tyr
	1595	1600	1605	
25	Leu Ser	Ala Asn Ser Gly Ala	Arg Leu Gly Ile Ala	Glu Glu Leu
	1610	1615	1620	
	Leu His	Ile Phe Lys Ala Ala	Phe Val Asp Pro Ala	Lys Pro Ser
	1625	1630	1635	
30	Met Gly	Ile Lys Tyr Leu Tyr	Leu Thr Pro Glu Thr	Leu Ser Thr
	1640	1645	1650	
35	Leu Ala	Lys Lys Gly Ser Ser	Val Thr Thr Glu Glu	Ile Glu Asp
	1655	1660	1665	
	Asp Gly	Glu Arg Arg His Lys	Ile Thr Ala Ile Ile	Gly Leu Ala
	1670	1675	1680	
40	Glu Gly	Leu Gly Val Glu Ser	Leu Arg Gly Ser Gly	Leu Ile Ala
	1685	1690	1695	

	Gly Ala	Thr Thr Arg Ala Tyr	Glu Glu Gly Ile Phe	Thr Ile Ser
	1700	1705	1710	
5	Leu Val	Thr Ala Arg Ser Val	Gly Ile Gly Ala Tyr	Leu Val Arg
	1715	1720	1725	
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	1730	1735	1740	
10	Thr Gly	Ala Gln Ser Leu Asn	Lys Val Leu Gly Arg	Glu Val Tyr
	1745	1750	1755	
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	1760	1765	1770	
15	Gly Thr	Thr His Leu Val Ala	Glu Ser Asp Leu Asp	Gly Ala Leu
	1775	1780	1785	
	Lys Val	Ile Gln Trp Leu Ser	Tyr Val Pro Glu Arg	Lys Gly Lys
20	1790	1795	1800	
	Ala Ile	Pro Ile Trp Pro Ser	Glu Asp Pro Trp Asp	Arg Thr Val
	1805	1810	1815	
25	Thr Tyr	Glu Pro Pro Arg Gly	Pro Tyr Asp Pro Arg	Trp Leu Leu
	1820	1825	1830	
	Glu Gly	Lys Pro Asp Glu Gly	Leu Thr Gly Leu Phe	Asp Lys Gly
	1835	1840	1845	
30	Ser Phe	Met Glu Thr Leu Gly	Asp Trp Ala Lys Thr	Ile Val Thr
	1850	1855	1860	
35	Gly Arg	Ala Arg Leu Gly Gly	Ile Pro Met Gly Val	Ile Ala Val
	1865	1870	1875	
	Glu Thr	Arg Thr Thr Glu Lys	Ile Ile Ala Ala Asp	Pro Ala Asn
	1880	1885	1890	
40	Pro Ala	Ala Phe Glu Gln Lys	Ile Met Glu Ala Gly	Gln Val Trp
	1895	1900	1905	

	Asn Pro	Asn Ala Ala Tyr Lys	Thr Ala Gln Ser Ile	Phe Asp Ile
	1910		1915	1920
5	Asn Lys	Glu Gly Leu Pro Leu	Met Ile Leu Ala Asn	Ile Arg Gly
	1925		1930	1935
	Phe Ser	Gly Gly Gln Gly Asp	Met Phe Asp Ala Ile	Leu Lys Gln
	1940		1945	1950
10	Gly Ser	Lys Ile Val Asp Gly	Leu Ser Asn Phe Lys	Gln Pro Val
	1955		1960	1965
	Phe Val	Tyr Val Val Pro Asn	Gly Glu Leu Arg Gly	Gly Ala Trp
	1970		1975	1980
15	Val Val	Leu Asp Pro Thr Ile	Asn Leu Ala Lys Met	Glu Met Tyr
	1985		1990	1995
	Ala Asp	Glu Thr Ala Arg Gly	Gly Ile Leu Glu Pro	Glu Gly Ile
20	2000		2005	2010
	Val Glu	Ile Lys Phe Arg Arg	Asp Lys Val Ile Ala	Thr Met Glu
	2015		2020	2025
25	Arg Leu	Asp Glu Thr Tyr Ala	Ser Leu Lys Ala Ala	Ser Asn Asp
	2030		2035	2040
	Ser Thr	Lys Ser Ala Glu Glu	Arg Ala Lys Ser Ala	Glu Leu Leu
	2045		2050	2055
30				
	Lys Ala	Arg Glu Thr Leu Leu	Gln Pro Thr Tyr Leu	Gln Ile Ala
	2060		2065	2070
35	His Leu	Tyr Ala Asp Leu His	Asp Arg Val Gly Arg	Met Glu Ala
	2075		2080	2085
	Lys Gly	Cys Ala Lys Arg Ala	Val Trp Ala Glu Ala	Arg Arg Phe
	2090		2095	2100
40				
	Phe Tyr	Trp Arg Leu Arg Arg	Arg Leu Asn Asp Glu	His Ile Leu
	2105		2110	2115

	Ser Lys	Phe Ala Ala Ala Asn	Pro Asp Leu Thr Leu	Glu Glu Arg
	2120	2125	2130	
5	Gln Asn	Ile Leu Asp Ser Val	Val Gln Thr Asp Leu	Thr Asp Asp
	2135	2140	2145	
	Arg Ala	Thr Ala Glu Trp Ile	Glu Gln Ser Ala Glu	Glu Ile Ala
	2150	2155	2160	
10	Ala Ala	Val Ala Glu Val Arg	Ser Thr Tyr Val Ser	Asn Lys Ile
	2165	2170	2175	
	Ile Ser	Phe Ala Glu Thr Glu	Arg Ala Gly Ala Leu	Gln Gly Leu
	2180	2185	2190	
15	Val Ala	Val Leu Ser Thr Leu	Asn Ala Glu Asp Lys	Lys Ala Leu
	2195	2200	2205	
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	<220>			
	<221>	misc_feature		
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	<223>	n is a, c, g or t		
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- 88 -

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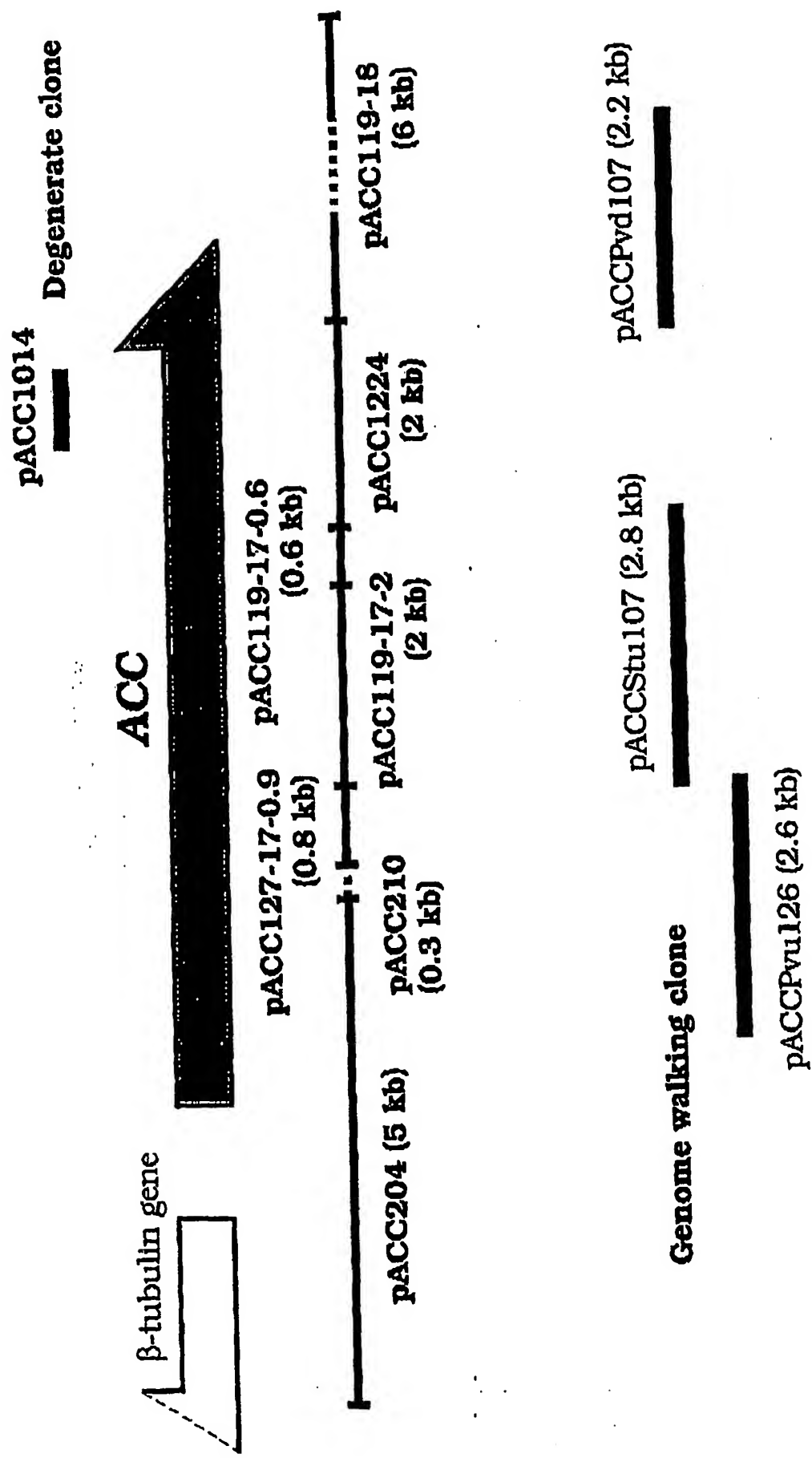


FIG.2 Cloning of ACC gene region from *P. rhodozyma*